



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5: C12N 15/62, C07K 15/00, C12P 21/00

A2

(11) International Publication Number:

WO 94/16085

(43) International Publication Date:

21 July 1994 (21.07.94)

(21) International Application Number:

PCT/US93/12687

(22) International Filing Date:

30 December 1993 (30.12.93)

(30) Priority Data:

07/998,271

30 December 1992 (30.12.92)

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(81) Designated States: CA, JP, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

Published

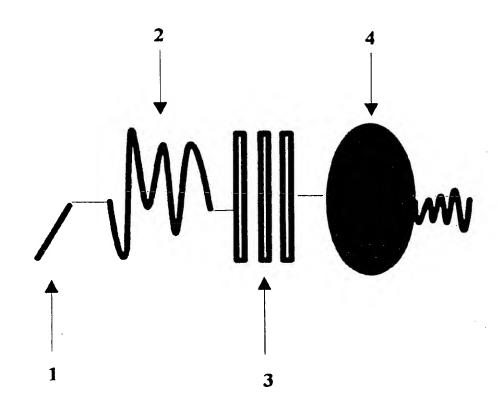
Without international search report and to be republished

upon receipt of that report.

(54) Title: HYBRID PROTEINS HAVING CROSS-LINKING AND TISSUE-BINDING ACTIVITIES

(57) Abstract

Hybrid proteins having crosslinking and tissue-binding activities, DNA molecules encoding such proteins and methods for producing the hybrid proteins from recombinant host cells are disclosed. The hybrid proteins disclosed herein are useful in tissue sealant and wound healing formulations.



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Description

Hybrid Proteins Having Cross-Linking and Tissue-Binding Activities

5 Technical Field

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The present invention relates generally toward methods for producing recombinant hybrid proteins, and more specifically, to methods for producing hybrid proteins from host cells through the use of recombinant DNA techniques.

Background of the Invention

The utilization of tissue sealants to replace or augment the use of mechanical wound closure devices has expanded in recent years in many surgical and trauma Tissue sealants include biological applications. adhesives (e.g. fibrin-based adhesives) and synthetic preparations (e.g. cyanoacrylates). Ιt is widely acknowledged that the use of synthetic preparations of tissue sealants is limited due to their toxicity and limited applications. Biological tissue adhesives have demonstrated utility in cases where the use of mechanical devices to close wounds is insufficient, such as joining blood vessels, closing holes in the dura, and in surgery on small or delicate tissues such as in the eye or ear.

biological tissue adhesives Fibrin-based generally contain fibrinogen, factor XIII and thrombin as principal ingredients, although in practice biological tissue adhesives are derived from whole blood and contain additional blood proteins. The fibrinogen and factor XIII components of these adhesives are prepared from pooled human plasma by cryoprecipitation (e.g. U.S. Patents No. 4,377,572; 4,362,567; 4,909,251), by ethanol precipitation (e.g. U.S. Patent No. 4,442,655) or from single donor plasma (e.g. U.S. Patent No. 4,627,879; Spotnitz et al., 166-168, 1989). The resultant Am. Surg. 55:

fibrinogen/factor XIII preparation is mixed with bovine thrombin immediately before use to convert the fibrinogen to fibrin and activate the factor XIII, thus initiating coagulation of the adhesive.

Fibrin-based tissue adhesives, in their current significant drawbacks that form, include standardization, lack of quality control from batch and the possibility of transmission of immunodeficiency virus (HIV), hepatitis virus and other etiologic agents. While recombinant production thrombin and factor XIII have been reported, proteins might be used in biological adhesives, the biological tissue adhesives still rely on large amounts of fibrinogen that is obtained from pooled blood. present, current fibrin(ogen)-based Αt tissue adhesives are not approved for use in the United States.

There is therefore a need in the art for tissue adhesive components, particularly components facilitate cross-linking to improve clot strength, are prepared at high levels with reproducible activity levels and which do not carry the possibility transmission of viral or other etiologic agents. The present invention addresses these needs by providing recombinant hybrid proteins that provide cross-linking and tissue-adhesive properties and that may be prepared at high levels.

Disclosure of the Invention

Briefly stated, the present invention provides hybrid proteins having cross-linking and tissue-binding activities, DNA molecules encoding such hybrid proteins and methods for producing hybrid proteins by recombinant means. In one aspect, In one aspect of the invention, the hybrid proteins comprise a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein. Within a related aspect of the

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invention, the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin. Within a preferred embodiment, the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926. Within another related aspect of the invention, the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin, the ten amino acid repeat beginning with glutamine amino acid number 496 of involucrin or the 400 amino-terminal amino acids of the fibrinogen α chain. Within a preferred embodiment of the invention, the tissue-binding domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336. Within a particularly preferred embodiment, the hybrid protein comprises the amino acid sequence of Sequence ID No. 6 from alanine, amino acid number 2 to proline, amino acid number 1336.

The present invention provides DNA molecules hvbrid proteins of the present invention comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein. embodiment, the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780. In another embodiment, the second DNA segment comprises the nucleotide sequence of Sequence ID 5 from nucleotide 2784 to nucleotide 4013. preferred embodiment, the DNA molecule comprises nucleotide sequence of Sequence ID Number from nucleotide 3 to nucleotide 4013.

In related embodiments of the invention, DNA constructs are provided which comprise a DNA molecule encoding a hybrid protein, whereins said DNA molecule

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comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule. Other embodiments of the invention concern host cells containing the DNA constructs of the present invention and methods of producing hybrid proteins.

10 Brief Description of the Drawings

Figure 1 discloses a representative hybrid protein containing (1) an N-terminal end-to-end interchain cross-linking domain, (2) a domain that promotes inter-chain cross-linking; (3) a domain that confers tissue binding activity; and (4) a carboxy-terminal domain that promotes end-to-end inter-chain cross-linking.

Figures 2-5 disclose absorbance time courses of representative cross-linking assays carried out in the presence of varying levels of factor XIII (activated to factor XIIIa via thrombin during the assay) or factor XIIIa.

Detailed Description of the Invention

The present invention provides novel hybrid cross-linking and tissue adhesive proteins having The hybrid proteins comprise a cross-linking activities. domain from a first protein covalently linked to a tissuebinding domain from a second protein. The hybrid proteins of the present invention are capable of cross-linking to themselves and to other proteins such as fibrin and fibrinogen and are capable of adhering to cell surfaces and/or extracellular matrix components. While not wishing to be bound by a graphical representation, Figure 1 shows a representative hybrid protein containing an N-terminal end-to-end inter-chain cross-linking domain; a domain that promotes inter-chain cross-linking; a domain that confers tissue binding activity; and a carboxy-terminal domain

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that promotes end-to-end inter-chain cross-linking. As used herein, cross-linking refers to the formation of covalent bonds between polypeptides.

The hybrid proteins of the present invention are useful as components of tissue sealant formulations to provide matrix material and to improve clot strength over a wound site, and as components in formulations that promote wound healing. The proteins of the present invention may contain native (i.e. wild-type) protein domains as well as domains that are allelic variants and genetically engineered or synthetic variants of the respective naturally occurring domains. Such variants are characterized by the presence of conservative amino acid substitutions and/or other minor additions, substitutions or deletions of amino acids.

As used within the context of the present invention, tissue-binding domains include protein domains containing amino acid sequences that facilitate adherence to cell surfaces and/or to extracellular matrix components fibronectin, hyaluronic as collagen, Fibronectin, for example, contains glycosaminoglycans. the sequence Gly-Arg-Gly-Asp-Ser (from amino acid 1614 through amino acid 1618 of Sequence I.D. No. 3) that has been shown to be central to cell recognition fibronectin receptor (for review see Yamada, Current Opinion in Cell_Biology 1: 956-963, 1989). The heparin binding domains of fibronectin (Sekiguchi et al., Proc. Natl. Acad. Sci. USA <u> 77:</u> 2661-2665, 1980), thrombospondin (Zardi et al., EMBO J. 6: 2337-3342, 1987 and Gutman and Kornblihtt, Proc. Natl. Acad. Sci. USA 84: 7179-7182, 1987) contain sequences that recognize heparin glycosaminoglycans sulfate-containing which extracellular matrix components. The collagen binding (Sekiguchi et al. 1980) domain of fibronectin ibid., contains amino acid sequences that bind the extracellular matrix component collagen.

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Particularly preferred tissue-binding domains are the heparin binding domain of fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 2 from alanine, amino acid number 1812 to valine, amino acid number 2171; the collagen binding domain of fibronectin. comprising the sequence of amino acids of Sequence I.D. No. 2 from glycine, amino acid number 282 to serine, amino acid number 608; and the amino terminal 229 amino acids of In this regard, a particularly preferred thrombospondin. tissue-binding domain is the cell-binding domain fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 3 from alanine, amino acid number 1357 to glutamic acid, amino acid number 1903. It will be evident to one skilled in the art that smaller portions of the cell-binding domain of fibronectin may be used within hybrid proteins of the present invention, particularly the sequence of amino acids of Sequence I.D. 3 from isoleucine, number 1532 through threonine, As noted above, it is generally amino acid number 1631. accepted that the sequence Gly-Arg-Gly-Asp-Ser acids 1614 to 1618 of Sequence I.D. No. 3) is central to cell recognition by fibronectin.

Cross-linking domains suitable for use in the hybrid proteins of the present invention are domains which contain amino acid sequences required for the formation of specific covalent bonds between peptide chains. In a preferred embodiment the inter-chain crosslinks are covalent bonds formed by the action of transglutaminase such as factor XIII, transglutaminase, prostate transglutaminase, keratinocyte transglutaminase, epidermal transglutaminase or placental transglutaminase. Transglutaminases catalyze formation of ϵ -(γ -glutamyl) lysine bonds between specific glutamine and lysine residues. However, other inter-chain cross-links, such as those formed by disulfide bonds, are also suitable cross-links. Suitable cross-linking domains include domains from the fibrinogen chain, α

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glutamine/lysine rich domains of loricrin that are involved in isodipeptide cross-link formation (Hohl al., J. Biol. Chem. 266: 6626-6636, 1991), and at least one of the 10 amino acid-long repeats of involucrin (Cell 46: 583-589, 1986 and Etoh et al., Biochem. Biophys. Res. Comm. 136: 51-56, 1986). Preferred cross-linking domains are the carboxy-terminal 103 amino acids of loricrin (Hohl et al., ibid.) and the ten-amino acid repeat beginning with glutamine, amino acid number 496 of involucrin (Simon (J. Biol. Chem. 263: 18093-18098, particularly preferred cross-linking domain comprises the 400 amino-terminal amino acids of the fibrinogen α chain (Doolittle et al., <u>Nature</u> 280: 464-468, 1979; Rixon et 22: 3250-3256, 1983). Biochemistry particularly, the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336 is preferred.

Although the hybrid proteins of the present invention may consist essentially of covalently linked cross-linking and tissue binding domains, they may further contain domains that facilitate end-to-end covalent cross-The γ chain of fibrinogen contains a domain that facilitates end-to-end cross-linking to another γ chain via ϵ -(γ -glutamyl)lysine bonds. This domain includes at acids 19 carboxy-terminal amino and more least the preferably includes the amino-terminal 275 amino acids of the fibrinogen γ chain. The α chain of fibrinogen contains an amino-terminal domain that is involved in interchain disulfide bond formation between α chains. This domain includes the amino-terminal portion of the α chain of fibrinogen from glycine, amino acid 36 to glycine, amino acid 67 of Sequence ID Number 4.

As will be evident to one skilled in the art, the hybrid proteins of the present invention may contain domains of human and other animal proteins. Proteins containing domains suitable for use in the present invention from human and other animals and the DNA

molecules encoding such proteins have been reported. fibrinogen and fibronectin, loricrin, Involucrin, example, have been studied in a variety of animals. DNA sequences encoding primate, canine and porcine involucrin have been reported (Djian and Green, Mol. Biol. Evol. 9: 417-432, 1992; Djian and Green, Proc. Natl. Acad. Sci. USA 88: 5321-5325, 1991 and Tseng and Green, Mol. Biol. Evol. 1103-1112, Mehrel et al. (Cell 61: 7: 293-302, 1990). have reported а DNA sequence encoding 1990) DNA sequences encoding rat and frog fibrinogen gamma chain have been reported (Haidaris and Courtney, Blood 79: 1218-1224, 1992 and Bhattacharya et al., Mol. <u>Cell. Endocrinol.</u> <u>72</u>: 213-220, 1990; respectively). sequences encoding chicken and lamprey fibrinogen α chains have been reported by Weissbach and Greininger (Proc. Natl. Acad. Sci. USA 87: 5198-5202, 1990) and Pan and Doolittle (Proc. Natl. Acad. Sci. USA 89: 2066-2070, 1992), respectively. DNA sequences encoding bovine and rat fibronectin have been reported by Petersen et (Proc. Natl. Acad. Sci. USA 80: 137-141, 1983) and Schwarzbauer et al., (Cell 35: 421-431, 1983). In general, it is preferred to prepare proteins that contain component domains from a single species to minimize the immunogenicity. the possibility of Thus, invention provides hybrid proteins that can be used in human and veterinary medicine.

invention According to the present having cross-linking and tissue adhesive proteins activities are produced recombinantly from host transformed with a DNA construct comprising a DNA segment encoding a cross-linking domain from a first joined to a DNA segment encoding a tissue-binding domain from a second protein. As used within the context of the present invention, two or more DNA coding sequences are said to be joined when, as a result of in-frame fusions between the DNA coding sequences or as a result of the of intervening sequences by normal cellular removal

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processing, the DNA coding sequences can be translated into a polypeptide fusion. Unless otherwise noted, the DNA segments may be joined in any order to result in a DNA coding sequence that can be translated into a polypeptide chain. Thus, the DNA segment encoding the tissue-binding domain may be joined to the 5' or the 3' end of the DNA segment encoding the cross-linking domain. However, as will be evident to one skilled in the art, the production of hybrid proteins that additionally include domains that facilitate end-to-end cross-linking will require that the DNA segments encoding such domains be positioned at the 5' and 3' termini of the molecules.

the present invention also isolated DNA molecules encoding hybrid proteins comprising a cross-linking domain from a first protein covalently linked to a tissue-binding domain from a second protein. In general, cDNA sequences are preferred for carrying out the present invention due to their lack of intervening sequences which can lead to aberrant RNA processing and reduced expression levels. DNA molecules encoding human fibronectin (Dufour et al., Exper. Cell Res. 193: 331-338, 1991) and a human fibrinogen α chain (Rixon et al., Biochemistry 22: 3250-3256, 1983) may be obtained from libraries prepared from liver cells according to standard laboratory procedures. It will be understood however, that suitable DNA sequences can also be obtained from genomic clones or can be synthesized de novo according to conventional procedures. If partial clones are obtained, it is necessary to join them in proper reading frame to produce a full length clone, using such techniques as endonuclease cleavage, ligation, and loop-out mutagenesis.

DNA sequences encoding hybrid proteins of the present invention may be prepared from cloned DNAs using conventional procedures of endonuclease cleavage, exonuclease digestion, ligation and in vitro mutagenesis. Alternatively, DNA sequences encoding the cross-linking

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and tissue-binding domains, such as those mentioned above, may be synthesized using standard laboratory techniques.

An exemplary DNA molecule encoding a hybrid protein having cross-linking and tissue-binding activities may be prepared by joining a DNA segment encoding at least the cell-binding domain of fibronectin and a DNA segment encoding at least an inter-chain cross-linking domain of fibrinogen at convenient restriction а site synthetic adapters to facilitate in-frame joining of the DNA segments. Alternatively, such DNA segments encoding hybrid proteins of the present invention may be prepared by joining the two domains at a convenient restriction site followed by loop-out mutagenesis to precisely remove unnecessary sequences and directly join the DNA segment encoding the cell-binding domain of fibronectin with the DNA segment encoding the cross-linking domain of fibrinogen.

DNA segments encoding the hybrid proteins of the instant invention are inserted into DNA constructs. used within the context of the present invention, a DNA construct is understood to refer to a DNA molecule, or a clone of such a molecule, either single- or doublestranded, which has been modified through human intervention to contain segments of DNA combined juxtaposed in a manner that would not otherwise exist in DNA constructs of the present invention comprise nature. a first DNA segment encoding a hybrid protein operably to additional DNA segments required expression of the first DNA segment. Within the context of the present invention, additional DNA segments will generally include promoters and transcription terminators, and may further include enhancers and other elements.

DNA constructs may also contain DNA segments necessary to direct the secretion of a polypeptide or protein of interest. Such DNA segments may include at least one secretory signal sequence. Secretory signal sequences, also called leader sequences, prepro sequences

and/or pre sequences, are amino acid sequences that act to direct the secretion of mature polypeptides or proteins from a cell. Such sequences are characterized by a core of hydrophobic amino acids and are typically (but not found at the amino termini of exclusively) segments encoding secretory synthesized proteins. DNA signal sequences are placed in-frame and in the correct spatial relationship to the DNA segment encoding the protein of interest in order to direct the secretion of the protein. Very often the secretory peptide is cleaved from the mature protein during secretion. Such secretory peptides contain processing sites that allow cleavage of the secretory peptides from the mature proteins as they the secretory pathway. Α preferred pass through processing site is a dibasic cleavage site, such as that recognized by the Saccharomyces cerevisiae KEX2 gene. particularly preferred processing site is a Lys-Arg processing site. Processing sites may be encoded within the secretory peptide or may be added to the peptide by, for example, in vitro mutagenesis.

Preferred secretory signals include the α factor signal sequence (pre-pro sequence: Kurjan and Herskowitz, Cell 30: 933-943, 1982; Kurjan et al., U.S. Patent No. 4,546,082; Brake, U.S. Patent No. 4,870,008), the PHO5 signal sequence (Beck et al., WO 86/00637), the BAR1 secretory signal sequence (MacKay et al., U.S. Patent No. 4,613,572; MacKay, WO 87/002670), the SUC2 signal sequence (Carlsen et al., Molecular and Cellular Biology 3: 439-447, 1983). Alternately, a secretory signal sequence may be synthesized according to the rules established, for example, by von Heinje (European Journal of Biochemistry 133: 17-21, 1983; Journal of Molecular Biology 184: 99-105, 1985; Nucleic Acids Research 14: 4683-4690, 1986).

secretory signal sequences may be used singly or may be combined. For example, a DNA segment encoding a first secretory signal sequence may be used in combination with a DNA segment encoding the third domain of barrier

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(described in U.S. Patent No. 5,037,243, which is incorporated by reference herein in its entirety). The DNA segment encoding the third domain of barrier may be positioned in proper reading frame 3' of the DNA segment of interest or 5' to the DNA segment and in proper reading frame with both the DNA segment encoding the secretory signal sequence and the DNA segment of interest.

The choice of suitable promoters, terminators and secretory signals is well within the level of ordinary skill in the art. Methods for expressing cloned genes in Saccharomyces cerevisiae are generally known in the art (see, "Gene Expression Technology," Methods in Enzymology, Vol. 185, Goeddel (ed.), Academic Press, San Diego, CA, 1990 and "Guide to Yeast Genetics and Molecular Biology," Methods in Enzymology, Guthrie and Fink (eds.), Academic Press, San Diego, CA, 1991; which are incorporated herein by reference). Transformation systems for other yeasts, including Hansenula polymorpha, Schizosaccharomyces pombe, Kluyveromyces lactis, Kluyveromyces fragilis, <u>Ustilago</u> maydis, Pichia pastoris, Pichia quillermondil and Candida maltosa are known in the art. See, for example, Gleeson et al., <u>J. Gen. Microbiol.</u> 132:3459-3465, 1986 and Cregg, U.S. Patent No. 4,882,279.

Proteins of the present invention can also be expressed in filamentous fungi, for example, strains of the fungi Aspergillus (McKnight et al., U.S. Patent No. 4,935,349, which is incorporated herein by reference). Methods for transforming <u>Acremonium chrysogenum</u> are disclosed by Sumino et al., U.S. Patent No. 5,162,228, which is incorporated herein by reference.

Other higher eukaryotic cells may also be used as hosts, including insect cells, plant cells and avian cells. Transformation of insect cells and production of foreign proteins therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and Bang et al., U.S. Patent No. 4,775,624, which are incorporated herein by reference. The use of <u>Agrobacterium rhizogenes</u> as a vector for

expressing genes in plant cells has been reviewed by Sinkar et al., J. Biosci. (Bangalore) 11:47-58, 1987.

Expression of cloned genes in cultured mammalian cells and in <u>E</u>. <u>coli</u>, for example, is discussed in detail in Sambrook et al. (<u>Molecular Cloning</u>: <u>A Laboratory Manual</u>, Second Edition, Cold Spring Harbor, NY, 1989; which is incorporated herein by reference). In addition to <u>E</u>. <u>coli</u>, <u>Bacillus</u> and other genera are useful prokaryotic hosts for expressing foreign proteins. As would be evident to one skilled in the art, one could express the proteins of the instant invention in other host cells such as avian, insect and plant cells using regulatory sequences, vectors and methods well established in the literature.

yeast, suitable vectors for use in the 15 In present invention include YRp7 (Struhl et al., Proc. Natl. Acad. Sci. USA 76: 1035-1039, 1978), YEp13 (Broach et al., <u>Gene</u> 8: 121-133, 1979), POT vectors (Kawasaki et al, U.S. Patent No. 4,931,373, which is incorporated by reference 20 herein), pJDB249 and pJDB219 (Beggs, Nature 275:104-108, 1978) and derivatives thereof. Preferred promoters for use in yeast include promoters from yeast glycolytic genes (Hitzeman et al., <u>J. Biol. Chem.</u> <u>255</u>: 12073-12080, 1980; Alber and Kawasaki, J. Mol. Appl. Genet. 1: 419-434, 1982; No. 4,599,311) Patent 25 Kawasaki, U.S. or dehydrogenase genes (Young et al., in Genetic Engineering of Microorganisms for Chemicals, Hollaender (eds.), p. 355, Plenum, New York, 1982; Ammerer, Meth. 192-201, 1983). In this 101: regard, Enzymol. particularly preferred promoters are the TPI1 promoter 30 . (Kawasaki, U.S. Patent No. 4,599,311, 1986) and the ADH2- 4^{\square} promoter (Russell et al., Nature 304: 652-654, 1983; Irani and Kilgore, U.S. Patent Application Serial No. 07/631,763, CA 1,304,020 and EP 284 044, which 35 incorporated herein by reference). The expression units include a transcriptional terminator. mav

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preferred transcriptional terminator is the <u>TPI1</u> terminator (Alber and Kawasaki, ibid.).

Host cells containing DNA constructs of present invention are then cultured to produce the hybrid The cells are cultured according to standard methods in a culture medium containing nutrients required for growth of the particular host cells. A variety of suitable media are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins, minerals and growth factors. The growth medium generally select for cells containing construct by, for example, drug selection or deficiency in which is complemented essential nutrient by selectable marker on the DNA construct or co-transfected with the DNA construct.

Selection of a medium appropriate particular host cell used is within the level of ordinary Yeast cells, for example, skill in the art. cultured in a chemically defined preferably comprising a non-amino acid nitrogen source, inorganic salts, vitamins and essential amino acid supplements. pH of the medium is preferably maintained at a pH greater than 2 and less than 8, preferably at pH 6.5. Methods for maintaining a stable pH include buffering and constant pH through the addition of preferably ammonium hydroxide. Preferred buffering hydroxide or agents include succinic acid and Bis-Tris (Sigma Chemical Co., St. Louis, MO). Yeast cells having a defect in a required for asparagine-linked glycosylation in a medium containing an preferably grown A preferred osmotic stabilizer is sorbitol stabilizer. supplemented into the medium at a concentration between 0.1 M and 1.5 M, preferably at 0.5 M or 1.0 M. Cultured mammalian cells are generally cultured in commercially available serum-containing or serum-free media.

The recombinant hybrid proteins expressed using the methods described herein are isolated and purified by

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conventional procedures, including separating the cells medium by centrifugation or filtration, from the the proteinaceous components of precipitating supernatant or filtrate by means of a salt, e.g. ammonium sulfate, purification by a variety of chromatographic procedures, e.g. ion exchange chromatography or affinity Methods the like. of chromatography, or protein purification are known in the art (see generally, Scopes, R., Protein Purification, Springer-Verlag, NY (1982),which is incorporated herein by reference) and may be applied to the purification of the recombinant proteins of the present invention.

The hybrid proteins of the present invention may as components of tissue adhesives. is be used preferred that the tissue adhesives be formulated to provide a concentration of the hybrid proteins of the present invention of between about 5 mg/ml to 100 mg/ml, with concentrations in the range of 35 to 50 mg/ml being particularly preferred. As disclosed above, tissue adhesives generally contain factor XIII and thrombin. Additional components may also be included in the tissue adhesive formulations. These additional components include growth factors such as PDGF, bFGF, TGFa, or EGF and protease inhibitors, such as aprotinin, transexamic acid, alpha-2 plasmin inhibitor, alpha-1-antitrypsin or the Pittsburgh mutant of alpha-1-antitrypsin (Arg-358 The tissue adhesives may alpha-1-antitrypsin). contain salts, buffering agents, reducing agents, bulking agents, and solubility enhancers. Albumin, NaCl, CaCl2, phosphate buffers, for citrate and example, may included. Preferably, the tissue adhesives of the present invention are prepared as lyophilized powders, liquid concentrates of ready-to-use liquids. Lyophilized powders are preferred for ease of handling and storage.

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The following examples are offered by way of illustration and not by way of limitation.

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EXAMPLES

Example 1 -Subcloning and Modification of ADH2 Promoters

An <u>ADH2-4^C</u> promoter was constructed as described in co-pending U.S. Patent Application 07/631,763, CA 1,304,020 and EP 284 044, which are incorporated herein by reference. A DNA construct comprising the complete <u>ADH2-4^C</u> promoter mutagenized at the 3' end to place an Eco RI site in place of the translation start codon, designated p410-4^C (deposited with the American Type Culture Collection (12301 Parklawn Dr., Rockville, MD 20852) under accession number 68861) was used as the source of the ADH2-4^C promoter.

A PAP-I cDNA (U.S. Patent No. 4,937,324) was with the $ADH2-4^{C}$ promoter. Plasmid pAP1.7, ioined comprising the 1.7 kb cDNA in pUC18, was cut with Nco I and Bam HI, and the linearized plasmid was isolated through two rounds of gel purification. The promoter from p410-4° was joined to the 5' end of the PAP-I cDNA via an Eco RI-Nco I adapter. The 1.2 kb Bam HI-Eco RI promoter fragment from p410-4°, Eco RI-Nco I adapter I-Bam HI linearized pAP1.7 plasmid were and the Nco The resultant plasmid was designed pPR1. presence of the correct promoter fusion was confirmed by DNA sequencing.

A yeast expression vector comprising the $\underline{ADH2-4^C}$ promoter, the PAP-I cDNA and the $\underline{TPI1}$ terminator was constructed. Plasmid pZUC13 (comprising the \underline{S} . cerevisiae chromosomal $\underline{LEU2}$ gene and the origin of replication from \underline{S} . cerevisiae 2 micron plasmid inserted into pUC13 and constructed in a manner analogous to pZUC12, described in published EP 195,691, using the plasmid pMT212, which is described in published EP 163 529) was cut with Bam HI. Plasmid pPR1 was digested completely digested with Bam HI and partially digested with Sac I to isolate the 2.1 kb $\underline{ADH2-4^C}$ promoter-PAP-I cDNA fragment. Plasmid pTT1 (described in detail below) was digested with Sac I and

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Bam HI to isolate the 0.69 bp <u>TPI1</u> terminator fragment. The Bam HI-Sac I fragment from pPR1 and the Sac I-Bam HI fragment from pTT1 were ligated with the Bam HI-linearized pZUC13. A plasmid containing the expression unit was designated pZ3.

Example 2 - Subcloning of the TPI1 terminator

The yeast <u>TPI1</u> terminator fragment was obtained from plasmid p270 described by Murray and Kelly (U.S. Patent 4,766,073, which is incorporated by reference herein in its entirety). Plasmid p270 contains the <u>TPI1</u> terminator inserted as and Xba I-Bam HI fragment into YEp13. Alternatively, the <u>TPI1</u> terminator may be obtained from plasmid pM220 (deposited with American Type Culture Collection as an <u>E. coli</u> RR1 transformant under accession number 39853) by digesting the plasmid with Xba I, and Bam HI and purifying the <u>TPI1</u> terminator fragment (~700 bp).

The <u>TPI1</u> terminator was removed from plasmid p270 as a Xba I-Bam HI fragment. This fragment was cloned into pUC19 along with another fragment containing the <u>TPI1</u> promoter fused to the CAT (chloramphenicol acetyl transferase) gene to obtain a <u>TPI1</u> terminator fragment with an Eco RV end. The resultant plasmid was designated pCAT. The <u>TPI1</u> terminator was then cut from pCAT as an Eco RV-Bam HI fragment and cloned into pIC19H (Marsh et al., <u>Gene 32</u>:481-486, 1984) which had been cut with the same enzymes, to obtain pTT1 (disclosed in U.S. Patent No. 4,937,324, which is incorporated herein by reference).

30 Example 3 - <u>Construction of Yeast Vectors pDPOT and pRPOT</u>

Plasmid pDPOT was derived from plasmid pCPOT (ATCC No. 39685) by replacing the 750 bp Sph I-Bam HI fragment of pCPOT containing 2 micron and pBR322 sequences with a 186 bp Sph I-Bam HI fragment derived from the pBR322 tetracycline resistance gene.

Plasmid pRPOT was derived from plasmid pDPOT by replacing the Sph I-Bam HI fragment with a polylinker. Plasmid pDPOT was digested with Sph I and Bam HI to isolate the 10.8 kb fragment. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were designed to form an adapter with a Bam HI adhesive end and an Sph I adhesive end flanking Sma I, Sst I and Xho I restriction sites. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were kinased and annealed to form the Bam HI-Sph I adapter. The 10.8 kb pDPOT fragment was circularized by ligation with the ZC1551/ZC1552 adapter (Sequence ID Nos. 7 and 8). The resultant plasmid was termed pRPOT.

Example 4 - <u>Construction of a Fibrinogen: Fibronectin</u> Hybrid cDNA Expression Vector

A. Construction of pFN14A

DNA construct containing DNA а encoding the fibronectin cell-binding domain operably linked to the $ADH2-4^{C}$ promoter in plasmid pUC19 The fibronectin coding sequence was obtained constructed. from plasmid pFH103 (Dufour et al., Exper. Cell Res. 193: 331-338, 1991). Plasmid pFH103 was digested with Nco I and isolate the 4 kb fragment containing the fibronectin coding sequence. Oligonucleotides ZC2052 and ZC2053 (Sequence ID Nos. 9 and 10) were designed provide, upon annealing, an adapter containing a 5' Eco RI end, an internal Nco I site, a DNA segment encoding a methionine and amino acids 979 through 981 of Sequence ID Number 2 and a 3' Nco I adhesive end that destroys the Nco I site. Oligonucleotides ZC2052 ZC2053 (Sequence ID Nos. 9 and 10) were annealed ligated with the 4 kb Nco I-Xba I fibronectin fragment into Eco RI-Xba I linearized pUC19. The resultant plasmid was designated pFN4.

Plasmid pFN4 was digested with Hind III and Apa I to isolate the 3.3 kb fibronectin fragment. Oligonucleotides ZC2493 and ZC2491 (Sequence ID Nos. 12

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and 11) were designed to provide, when annealed, an Apa I-Xba I adapter encoding the amino acids Pro and Phe followed by a stop codon. The oligonucleotides were annealed and combined with the 3.3 kb Hind III-Apa I fragment and Hind III-Xba I linearized pUC19 to form plasmid pFN7. Plasmid pFN7 comprises a DNA segment encoding amino acids 1273-2186 of Sequence ID Number 2 followed by an in-frame stop codon.

The <u>ADH2-4</u>^C promoter was joined to the 5' end of the fibronectin cDNA in plasmid pFN5. Plasmid pFN4 was digested with Nco I and Hind III to isolate the 0.89 kb fibronectin coding sequence. Plasmid pZ3 (described in detail above) was digested with Bam HI and Nco I to isolate the 1.25 kb <u>ADH2-4</u>^C promoter fragment. The 1.25 kb Bam HI-Nco I promoter fragment and the Nco I-Hind III fibronectin coding sequence fragment were ligated to Bam HI-Hind III linearized pUC19 to form plasmid pFN5.

Plasmid pFN5 was digested with Bam HI and Hind III to isolate the 2.1 kb promoter-fibronectin fragment. Plasmid pFN7 was digested with Hind III and Xba I isolate the 2.8 kb fibronectin fragment that was modified to encode a stop codon following the Pro-Phe sequence. The TPI1 terminator sequence was obtained from pTT1 as a 0.7 kb Xba I-Sal I fragment. The 2.1 kb Bam HI-Hind III promoter-fibronectin fragment, the 2.8 kb Hind III-Xba I fibronectin fragment and the 0.7 kb TPI1 terminator fragment were joined in a four-part ligation with Bam HIlinearized pRPOT. Α plasmid containing the fibronectin expression unit in the pRPOT vector was designated pR1.

The original clone pFH103 contained a frame-shift mutation in the EIIIB region of the fibronectin cDNA. The mutation was corrected by the replacement of the region with an analogous region from the plasmid pFHA3 (obtained from Jean Paul Thiery, Laboratoire de Physiopathologie du Developpement, CNRS URA 1337, Ecole Normale Superiure, 46 rue d'Ulm, 75230 Paris Cedex 05,

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France). Plasmid pFHA3 was derived from pFH103 by excising the 3211 qd Xba I-Asp 718I fragment of fibronectin, blunting of the resultant adhesive ends and religating. Plasmid pFHA3 contains a DNA segment encoding the signal and propeptides, the first three and one half type I repeats, and the carboxy-terminal half of human fibronectin from the middle of the EIIIB segment.

Plasmid pR1 was digested with Bam HI and Kpn I isolate the 2.2 kb promoter-fibronectin fragment. Plasmid pFH Δ 3 was digested with Kpn I and Apa I to isolate the internal fibronectin fragment that corrects the frameshift mutation present in the parent cDNA from pFH103. Plasmid pR1 was digested with Apa I and Bam HI to isolate the TPI1 terminator fragment. The 2.2 kb Bam HI-Kpn I promoter-fibronectin fragment, the 2.75 kb Kpn I-Apa I internal fibronectin fragment and the 0.69 kb Apa I-Bam HI terminator fragment were joined in a four-part ligation with Bam HI-linearized pDPOT. The resulting construction was designated pD32.

A DNA segment encoding the <u>ADH2-4</u>^C promoter and initiation methionine from plasmid pD32 was subcloned into pIC19H (Marsh et al., <u>Gene 32</u>:481-486, 1984) as a 1.25 kb Bam HI-Nco I fragment. Plasmid pD32 was also digested with Nco I and Bgl II to isolate the 3 kb fibronectin cDNA fragment encoding amino acids 979-1972 of Sequence ID Number 2. The 1.25 kb Bam HI-Nco I fragment and the Nco I-Bgl II fragment were ligated with Bam HI-linearized pIC19H. A plasmid containing a Bam HI site proximal to the <u>ADH2-4</u>^C promoter was designated pFN14A.

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B. - Construction of Plasmid pD38

An expression vector comprising a DNA segment encoding a fibronectin-fibrinogen hybrid protein operably linked to the $\underline{ADH2-4}^{\underline{C}}$ promoter and the TPI1 terminator was constructed. To assemble the DNA sequence encoding the hybrid protein, a DNA segment encoding approximately the

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carboxy-terminal 409 amino acids of the α chain of fibrinogen was first subcloned.

A fibrinogen α chain cDNA was obtained from Dominic W. Chung (Department of Biochemistry, University of Washington, Seattle, WA) in plasmid pHI α 3 (Rixon et al., <u>Biochemistry 22</u>: 3250-3256, 1983). Sequence analysis of the cDNA insert in plasmid pHI α -3 revealed a deletion of codons 1348-1350 of the published sequence resulting in the deletion of Serine, amino acid 417.

The DNA segment encoding the carboxy-terminus of the fibrinogen α chain was subcloned into plasmid pUC19. Plasmid pHIa-3 was digested with Asp 718 and Ssp I to isolate the approximately 2 kb fragment encoding the carboxy-terminus of the fibrinogen α chain from amino acid 244 to amino acid 643 and some 3' untranslated sequence of Plasmid pTT1 was digested with Eco Sequence ID Number 4. RV and Sal I to isolate the approximately 700 bp TPI1 terminator fragment. The 2 kb fibrinogen α chain sequence and the TPI1 terminator sequence were ligated with pUC19 that had been linearized with Asp 718 and Sal I. ligation mixture was transformed into E. coli, and plasmid DNA was prepared and analyzed by restriction endonuclease and DNA sequence analysis. DNA sequence analysis of a candidate clone revealed that the Sal I site joining the TPI1 terminator sequence and the pUC19 polylinker site was Plasmid DNA from the candidate clone was not present. HI to liberate the digested with Asp 718 and Bam approximately 1.9 kb fibrinogen-TPI1 terminator fragment.

To join the fibronectin coding sequence with the fibrinogen α chain sequence, synthetic oligonucleotides were synthesized to provide, when annealed, a Sal I-Asp 718 adapter encoding an internal Afl II restriction site, and a sequence encoding amino acids 1886 through 1903 of fibronectin (Sequence ID Number 2), a glycine residue and amino acids 235 through 243 of the fibrinogen α chain (Sequence ID Number 4). Oligonucleotides ZC3521 and ZC3522 (Sequence ID Nos. 13 and 14) were annealed. The

1.9 kb Asp 718-Bam HI fibrinogen-TPI1 terminator fragment and the Sal I-Asp 718 ZC3521/ZC3522 adapter (Sequence ID Nos. 13 and 14) were ligated with pUC19 that had been linearized with Sal I and Bam HI. The resultant plasmid was designated pFG4.

The DNA segment encoding the fibronectinfibrinogen α chain sequence in plasmid pFG4 was joined encoding DNA segment the amino-terminal fibronectin sequence (from amino acid 989 to amino acid ID Number 2) 1885 of Sequence in plasmid construct plasmid pD37. Plasmid pFN14A was digested with Bgl II and Afl II to isolate the approximately 3.9 kb ADH2-4^C promoter-fibronectin fragment. Plasmid pFG4 was Afl digested with ΙI and Bam HI to isolate approximately 2 kb fibronectin-fibrinogen-TPI1 terminator The 3.9 kb Bgl II-Afl II fragment and the 2 kb Afl II-Bam HI fragment were ligated with Bam HI-linearized A plasmid with the expression unit inserted with the direction of transcription in the same direction as the POT1 gene in the pDPOT vector was designated pD37.

To place the expression unit present in pD37 in the opposite orientation, such that the direction transcription of the expression unit was in the opposite direction to that of the POT1 gene, plasmid pD37 was digested with Nco I and Xba I to isolate the approximately 4 kb fibronectin-fibrinogen α chain fragment. pFN14A was digested with Bam HI and Nco I to isolate the approximately 1.3 kb ADH2-4^C promoter fragment. pTT1 was digested with Bam HI and Xba I to isolate the approximately 700 bp TPI1 terminator fragment. HI-NCO I ADH2-4^C promoter fragment, the Nco I-Xba I fibronectin-fibrinogen α chain fragment and the Xba I-Bam TPI1 terminator fragment were ligated with Bam HIlinearized pDPOT that had been treated with calf alkaline phosphatase to prevent recircularization. containing the expression unit in the opposite orientation relative to the POT1 gene was designated pD38.

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nucleotide sequence and deduced amino acid sequence of the DNA segment encoding the fibronectin-fibrinogen hybrid of plasmid pD38 is shown in Sequence ID Number 5. Plasmid pD38 was deposited on December 15, 1992 with the American Type Culture Collection (12301 Parklawn Drive, Rockville, MD) as an <u>E. coli</u> transformant.

Example 5 - Expression of a Fibronectin-Fibrinogen Hybrid Protein in Yeast

transformed 10 Plasmid pD38 was into Saccharomyces cerevisiae host strain ZM118 $(\underline{MATa}/\underline{MAT}\alpha)$ ura3/ura3 Δtpi1::URA3/Δtpi1::URA3 leu2-3,112/leu2-3,112 bar1/bar1 pep4::URA3/pep4::URA3 [cir0]) using essentially the method described by Hinnen et al. (Proc. Natl. Acad. 1929-1933, 1978). Transformants were 15 <u> 75:</u> Sci. USA selected for their ability to grow on medium containing glucose as the sole carbon source.

The ZM118[pD38] transformant was scaled up in a liter fermenter to facilitate purification of the hybrid protein. A single ZM118[pD38] colony was selected from a YEPD + Ade + Leu plate (Table 1) and inoculated into -LeuTrpThrD medium (Table 1). The culture was incubated for approximately 52 hours after which the cells were harvested. The cells were washed in T.E. buffer (Sambrook et al., ibid.), resuspended in T.E. buffer + 30% glycerol, and aliquotted into 1 ml seed vials. seed vials were stored at -80°C. One seed vial was used to inoculate 100 ml of YEPD + Ade + Leu (Table 1). culture was grown for approximately 28 hours to a final The 100 ml culture of ZM118[pD38] was A660 of 7.7. inoculated into a 10 liter fermenter with a final volume of 6.0 liters of medium containing 10 g/L (NH₄)₂SO₄, 5 g/L KH_2PO_4 , 5 g/L $MgSO_4$: $7H_2O_4$, 1 g/L NaCl, 0.5 g/L $CaCl_2$: $2H_2O_4$ 3.68 q/L A.A.I. (Table 1), 4.2 g/L citric acid, 60 g/L glucose, 10 ml/L Trace Metal Solution (Table 1), 0.4 ml/L PPG-2025 (Polypropylene glycol, MW 2025, Union Carbide Corp, Danbury, CT) that had been pH adjusted to pH 5.0

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with NaOH. In addition to the inoculation culture, 30 ml of Vitamin solution was added (Table 1). The culture was grown for 23 hours at 30°C with the addition of 2 M NaOH to maintain pH of approximately 5.

Table 1 Media Recipes

5	-LeuThrTrp Amino Acid Mixture
	4 g adenine
	3 g L-arginine
	5 g L-aspartic acid
	2 g L-histidine free base
10	6 g L-isoleucine
	4 g L-lysine-mono hydrochloride
	2 g L-methionine
	6 g L-phenylalanine
	5 g L-serine
15	5 g L-tyrosine
	4 g uracil
	6 g L-valine
	Mix all the ingredients and grind with
20	a mortar and pestle until the mixture is finely
	ground.
	-LeuTrpThrD
	20 g glucose
25	6.7 g Yeast Nitrogen Base without amino
	acids (DIFCO Laboratories, Detroit,
	MI)
	0.6 g -LeuThrTrp Amino Acid Mixture
	18 g Agar
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	Mix all the ingredients in distilled
	water. Add distilled water to a final volume of
	1 liter. Autoclave 15 minutes. Pour plates and
	allow to colidify

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Table 1 continued

	YEPD + Ade + Leu Plates		
	20 g	glucose	
	20 g	Bacto Peptone (DIFCO Laboratories)	
5	10 g	Bacto Yeast Extract (DIFCO	
		Laboratories)	
	18 g	agar	
	4 ml	1% adenine	
	8 ml	1% L-leucine	
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		Mix all ingredients in distilled	
	water, ar	nd bring to a final volume of 1 liter.	
	Autoclave	25 minutes and pour plates.	
15	YEPD + Ad	e + Leu Medium	
	20 g	glucose	
	20 g	Bacto Peptone (DIFCO Laboratories)	
	10 g	Bacto Yeast Extract (DIFCO	
		Laboratories)	
20	4 ml	1% adenine	
	8 ml	1% L-leucine	
		Mix all ingredients in distilled	
	water, an	nd bring to a final volume of 1 liter.	
25	Autoclave	25 minutes.	

Table 1 continued

	A.A.I.	
	4.0 g	adenine
	5.0 g	L-alanine
5	2.0 g	L-arginine
	5.0 g	L-asparagine
	5.0 g	L-aspartic acid
	5.0 g	L-cysteine
	5.0 g	L-glutamine
10	5.0 g	L-glutamic acid
	5.0 g	L-glycine
	8.0 g	L-histidine
	5.0 g	L-isoleucine
	3.0 g	L-lysine-mono hydrochloride
15	2.0 g	L-methionine
	5.0 g	L-phenylalanine
	5.0 g	L-proline
	5.0 g	L-serine
	5.0 g	L-threonine
20	2.0 g	L-tryptophan
	3.0 g	L-tyrosine
	3.0 g	uracil
	5.0 g	L-valine
25		Mix all the ingredients and grind with
	a mortar	and pestle until the mixture is finely

ground. Store at room temperature.

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Table 1 continued

	Trace Metal Solution	
	0.68 g	ZnCl ₂
	5.4 g	FeCl ₃ .6H ₂ O
5	1.91 g	$MnCl_2\cdot 4H_2O$
	0.22 g	CuSO ₄ .5H ₂ O
	0.258 g	CoCl ₂
	0.062 g	H_3BO_3
	0.002 g	$(NH_4)_6Mo_2O_2$
10	0.002 g	KI
	10.0 ml 37	7% HCl

Dissolve solids in water and bring to a final volume of 1 liter.

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	25 mg	d-biotin
	400 mg	thiamine
	400 mg	pyridoxine
20	7.5 g	meso-inositol
	7.5 g	Ca pantothenate
	300 mg	niacinamide
	50 mg	folic acid
	100 mg	riboflavin

choline

500 mg

Vitamin Solution

Dissolve solids in water and bring to a final volume of 1 liter.

A 60 liter fermenter with a final volume of 50 liters of medium containing 60 g/L yeast extract (Universal Foods, Milwaukee, WI), 2.5 g/L MgSO₄·7H₂O (Mallinkrodt Inc., St. Louis, MO), 1 g/L CaCl₂·2H₂O (Mallinkrodt, Inc.), 1 g/L KCl (Mallinkrodt, Inc.), 10 ml/L of Trace Metal Solution (Table 1), 0.5 ml/L PPG-2025 (Union Carbide) that had been adjusted to a pH of 5.0 with

H₃PO₄ was prepared, and the medium was sterilized. sterilization, 5.0 liters of the 23 hour fermentation culture and 500 ml of Vitamin Solution (Table 1) were inoculated into the medium. During the fermentation, a solution of 50% glucose, 5% (NH₄)₂SO₄, 0.05% citric acid was fed into the fermenter at a rate of 150 ml/hour, pH was maintained at approximately pH 5 addition of 2 M NH4OH. PPG-2025 was added as needed to approximately 49 hours control foaming. Αt inoculation, an ethanol feed was begun by the addition of ethanol to the fermenter at a rate of 150 ml/min. culture was grown for a total of 67.25 hours at 30°C.

At the end of the fermentation, 50 liters of the culture was diluted to 100 liters with water. were removed from the spent medium by centrifuging 50 liters at a time through a Westfalia CSA 19 centrifuge (Westfalia, Oelde, Germany) at a flow rate liters/min. The cells were rinsed with water. From the centrifugation, approximately 20 liters of cell slurry containing approximately 35% cells was obtained. were added to the slurry to achieve a final concentration of the following salts: 50 mM NaCl, 10 mM Na2HPO4, 5 mM The cell slurry was passed through a Dynomill bead mill using 0.5 mm lead-free glass beads (Willy A Bachofen AG MashinenFabrik, Basle, Switzerland) at a rate of 4 The Dynomill was rinsed with Lysis liters per minute. buffer (50 mM NaCl, 10 mM Na $_2$ HPO $_4$, 5 mM EDTA, pH 7.2) to a final volume of 80 liters. The final slurry had a pH of approximately 10°C and a temperature of conductivity of 5 ms/cm.

The cell slurry was subjected to centrifugation as described above, and the cell pellet was rinsed with lysis buffer. After centrifugation approximately 20 liters of cell slurry was obtained. The cell slurry was extracted by first adjusting the concentration of the cell debris to approximately 40-50% with lysis buffer. Solid urea, NaCl and EDTA were added to the cell slurry to

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achieve a final concentration of approximately 8 M urea, NaCl and 10 mM EDTA. The approximate concentrations were obtained by the addition of 450 g/L of urea, 18 g/L of NaCl and 4.2 g/L of EDTA. The cell slurry was adjusted to pH 7.8 with 0.5 M NaOH. The solids were dissolved into the slurry and the pellets were extracted for a total of 50 minutes. Following extraction, the mixture was diluted 1 to 4 with water, adjusted to a conductivity of 12.5 ms/cm with NaCl and adjusted to a pH of 9.5 with 0.5 M NaOH.

The extracted slurry centrifuged was as described above with the lysis buffer rinse. The the supernatant was adjusted to pH 9.5 with 0.5 M NaOH. The supernatant was analyzed by SDS polyacrylamide gel electrophoresis (SDS-PAGE) using the PHAST Separation and Control Unit (Pharmacia LKB Biotechnology Inc., Piscataway, NJ), and the protein was visualized using Coomassie Blue staining. A 2 liter Q-sepharose column (Pharmacia) was equilibrated at 5 liters/hour with successive washes of the following solutions: 8 liters of 3 M urea, 1 M NaCl, 50 mM glycine, pH 11.5; 5 liters of 0.5 M NaOH; 1.5 liters of water; 5 liters of 0.1 M HCl; and 6.0 liters of Wash buffer (50 mM glycine, 90 mM NaCl, with a conductivity of 12.5 ms/cm). supernatant (110 liters) was then applied to the column at 5 liters per hour.

The column ran dry after loading the The gel was resuspended in Wash buffer and supernatant. The repacked column was washed with 4 liters of 50 mM glycine, 90 mM NaCl, 5 mM EDTA, pH 10.0. material was eluted with elution buffer (50 mM glycine, 5 mM EDTA (pH 9.9) with a final concentration of NaCl giving a conductivity of 30.2 cm/ms (approximately 270 mM NaCl)). at 100 ml per minute. The approximately 600 ml fractions were collected after the conductivity of the reached the conductivity of the elution buffer. Fractions

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were analyzed by SDS-PAGE analysis as described above and fractions 1 through 10 were pooled.

The pooled fractions were then applied to a 2 liter phenyl Sepharose column (Pharmacia) that had been equilibrated by successive washes at 5 liters per hour with the following solutions: 3 liters of 0.5 M NaOH; 3 liters of water; 3 liters of 2 M urea, 50 mM glycine, pH 10.5; 1.5 liters of water; 3 liters of 0.1 M HCl; and 3 liters of Equilibration buffer (50 mM glycine, 2.5 M NaCl, 2 mM EDTA (pH 10.0) with a conductivity of 180 ms/cm). The pooled peak fractions, which had been adjusted to a conductivity of 180 ms/cm with NaCl and a pH of 10.0 with 0.5 M NaOH, were loaded onto the phenyl sepharose column. Following the loading of the peak fractions, the column was washed with Equilibration buffer. The column was eluted with 6 liters of 50 mM glycine, 2 mM EDTA :(pH 10.25) with a NaCl concentration giving the solution a conductivity of 96 ms/cm. The conductivity of the eluant was measured throughout the elution. The conductivity of the eluant upon starting the elution was 180 ms/cm. : In the third fraction, the conductivity of the eluant dropped At this point, the elution buffer was 96 ms/cm. changed to a buffer having the conductivity of 42 ms/cm. The eluant was collected through fraction number 8.

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Example 6 - <u>Cross-Linking Assay Using the Hybrid</u> Fibringen-Fibronectin Protein

ability of the purified fibrinogen-The protein form fibronectin hybrid to transglutminasecatalyzed interchain cross links assessed. was transglutaminase activity was provided by the addition of recombinant factor XIII and thrombin or by the addition of recombinant factor XIIIa.

10 A. Preparation of Factor XIII

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Recombinant factor XIII was prepared essentially as described in co-pending U.S. Patent Application No. 07/927,196, which is incorporated by reference herein in its entirety. Briefly, factor XIII was isolated from a strain of the yeast Saccharomyces cerevisiae that had been transformed with an expression vector capable of directing the expression of factor XIII. The factor XIII-producing cells were harvested and lysed, and a cleared lysate was The lysate was fractionated by anion exchange prepared. chromatography at neutral to slightly alkaline pH using a column of derivatized agarose, such as DEAE FAST-FLOW SEPHAROSE (Pharmacia LKB Biotechnology, Piscataway, NJ) or the like. Factor XIII was then precipitated from the column eluate by concentrating the eluate and adjusting the pH to between 5.2 and 5.5, such as by diafiltration against ammonium succinate buffer. The precipitate was then dissolved and further purified using conventional chromatographic techniques, such as gel filtration hydrophobic interaction chromatography. The purified dialyzed, filtered, aliquotted factor XIII was factor XIIIa content was determined lyophilized. The (Bishop et al., Biochemistry 29: 1861-1869, 1990, which is incorporated by reference herein in its entirety) fluorometric assay of the dissolved, thrombin-activated material.

Factor XIII was activated to factor XIIIa by adding 2 U of thrombin per 100 mg of factor XIII. The

factor XIII was dissolved in buffer (20 mM sodium borate (pH 8.3), 1 mM $CaCl_2$). The thrombin was added, and the reaction was incubated at room temperature for twenty minutes.

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B. Cross-Linking Assays

The level of cross-linking between the hybrid proteins was measured as a rise in the absorbance at 350 nm over time in reaction mixtures containing the hybrid protein, factor XIII and thrombin or the hybrid protein 10 XIIIa. Control reactions were prepared factor containing factor XIII and thrombin or factor XIIIa alone. Cross-linking reactions were carried out in 1 ml cuvettes. For cross-linking reactions containing factor XIII thrombin, each reaction mixture was set up by placing 110 15 μ1 containing 40 Units of factor XIII, 36.7 μl containing 13 Units of factor XIII or 12.2 μ l containing 4 Units of factor XIII (described above) in one corner of the cuvette and 20 μ l containing 4 Units of thrombin (Sigma) in the opposite corner such that the solutions were not mixed. 20 The reaction was initiated by the addition of 1 ml of 2 mg/ml hybrid protein in buffer (10 mM Tris (pH 7.6), 20 mM sodium borate, 140 mM NaCl, 10 mM CaCl2). The absorbance of each reaction was read at 350 nm with the addition of protein being the first absorbance point. 25 linking reactions containing factor XIIIa, each reaction was set up by placing 110 μ l containing 40 Units of factor XIIIa, 36.7 μ l containing 13 Units of factor XIIIa or 12.2 µ1 containing 4 Units of factor XIIIa in the cuvette and adding 1 ml of 2 mg/ml hybrid in buffer (10 mM Tris (pH 30 7.6), 140 mM NaCl, 10 mM CaCl₂). The absorbance of the solution was read at 350 nm as described above. Analysis of the data generated from the absorbance time courses showed a sharp increase in absorbance in the presence of hybrid protein and the active transglutaminase relative to 35 the rise in absorbance in the absence of hybrid protein WO 94/16085 PCT/US93/12687

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(Figures 2-5). The results indicated that the hybrid protein is capable of transglutaminase-induced crosslinking.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviation from the spirit and scope of the invention. Accordingly, the invention is not to be limited except as by the following claims.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Irani, Meher H.
 - (ii) TITLE OF INVENTION: HYBRID CROSS-LINKING PROTEINS
 - (iii) NUMBER OF SEQUENCES: 14
 - (iv) CORRESPONDENCE ADDRESS:
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 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk

 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: WO
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 07/998,271
 - (B) FILING DATE: 31-DEC-1992
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- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7803 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS

(B) LOCATION: 6..7346

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1: TCAAC ATG CTT AGG GGT CCG GGG CCC GGG CTG CTG CTG GCC GTC 47 Met Leu Arg Gly Pro Gly Pro Gly Leu Leu Leu Ala Val 95 CTG TGC CTG GGG ACA GCG GTG CCC TCC ACG GGA GCC TCG AAG AGC AAG Leu Cys Leu Gly Thr Ala Val Pro Ser Thr Gly Ala Ser Lys Ser Lys 25 20 AGG CAG GCT CAG CAA ATG GTT CAG CCC CAG TCC CCG GTG GCT GTC AGT 143 Arg Gln Ala Gln Gln Met Val Gln Pro Gln Ser Pro Val Ala Val Ser 40 35 191 CAA AGC AAG CCC GGT TGT TAT GAC AAT GGA AAA CAC TAT CAG ATA AAT Gln Ser Lys Pro Gly Cys Tyr Asp Asn Gly Lys His Tyr Gln Ile Asn 55 CAA CAG TGG GAG CGG ACC TAC CTA GGT AAT GTG TTG GTT TGT ACT TGT 239 Gln Gln Trp Glu Arg Thr Tyr Leu Gly Asn Val Leu Val Cys Thr Cys 65 TAT GGA GGA AGC CGA GGT TTT AAC TGC GAA AGT AAA CCT GAA GCT GAA 287 Tyr Gly Gly Ser Arg Gly Phe Asn Cys Glu Ser Lys Pro Glu Ala Glu 80 85 GAG ACT TGC TTT GAC AAG TAC ACT GGG AAC ACT TAC CGA GTG GGT GAC 335 Glu Thr Cys Phe Asp Lys Tyr Thr Gly Asn Thr Tyr Arg Val Gly Asp 110 105 100 95 383 ACT TAT GAG CGT CCT AAA GAC TCC ATG ATC TGG GAC TGT ACC TGC ATC Thr Tyr Glu Arg Pro Lys Asp Ser Met Ile Trp Asp Cys Thr Cys Ile 120 115 GGG GCT GGG CGA GGG AGA ATA AGC TGT ACC ATC GCA AAC CGC TGC CAT 431 Gly Ala Gly Arg Gly Arg Ile Ser Cys Thr Ile Ala Asn Arg Cys His 130 135 GAA GGG GGT CAG TCC TAC AAG ATT GGT GAC ACC TGG AGG AGA CCA CAT 479 Glu Gly Gly Gln Ser Tyr Lys Ile Gly Asp Thr Trp Arg Arg Pro His 150 145 GAG ACT GGT GGT TAC ATG TTA GAG TGT GTG TGT CTT GGT AAT GGA AAA 527 Glu Thr Gly Gly Tyr Met Leu Glu Cys Val Cys Leu Gly Asn Gly Lys 165 160 GGA GAA TGG ACC TGC AAG CCC ATA GCT GAG AAG TGT TTT GAT CAT GCT 575 Gly Glu Trp Thr Cys Lys Pro Ile Ala Glu Lys Cys Phe Asp His Ala 190 175 623 GCT GGG ACT TCC TAT GTG GTC GGA GAA ACG TGG GAG AAG CCC TAC CAA Ala Gly Thr Ser Tyr Val Val Gly Glu Thr Trp Glu Lys Pro Tyr Gln 195 200 205

GG(Gl)	C TG	G AT p Me	G AT t Me 21	t Va	A GA 1 As	T TG p Cy:	T AC	T TG r Cy: 21	s Lei	G GG/ u Gly	A GA,	A GG u G1	C AG y Se 22	r Gl	A CGC y Arg	671
ATC Ile	ACT Thi	T TG Cy 22	s In	T TC r Se	T AG	A AAT g Asr	7 AG/ 1 Arg 230	g Cys	C AA(s Asr	C GAT	CA(G GA(n As) 23	Th:	A AG r Ar	G ACA g Thr	719
TCC Ser	TA1	^ Ar	A AT	T GG/ e Gl:	A GAO	C ACC D Thr 245	Tr	G AGO Ser	Lys	G AAG S Lys	GA7 Asp 250) Asr	r CG/ n Arg	A GG G G1	A AAC y Asn	767
CTG Leu 255	Let	CA(Glr	G TGO	C ATO	C TG(Cys 260	Thr	GGC Gly	AAC Asn	GGC Gly	CGA Arg 265	Gly	A GAG	TG(Trp	AA(G TGT S Cys 270	815
GAG Glu	AGG Arg	CAC His	C ACC Thr	TC1 Ser 275	· Val	CAG Gln	ACC Thr	ACA Thr	TCG Ser 280	Ser	GGA Gly	TCT Ser	GGC Gly	CC(Pro 285	TTC Phe	863
ACC Thr	GAT Asp	GTT Val	CGT Arg 290	Ala	GCT Ala	GTT Val	TAC Tyr	CAA Gln 295	Pro	CAG Gln	CCT Pro	CAC His	CCC Pro 300	Glr	CCT Pro	911
CCT Pro	CCC Pro	TAT Tyr 305	ыу	CAC His	TGT Cys	GTC Val	ACA Thr 310	GAC Asp	AGT Ser	GGT Gly	GTG Val	GTC Val 315	TAC Tyr	TCT Ser	GTG Val	959
GGG Gly	ATG Met 320	CAG Gln	TGG Trp	TTG Leu	AAG Lys	ACA Thr 325	CAA Gln	GGA Gly	AAT Asn	AAG Lys	CAA Gln 330	ATG Met	CTT Leu	TGC Cys	ACG Thr	1007
TGC Cys 335	CTG Leu	GGC Gly	AAC Asn	GGA Gly	GTC Val 340	AGC Ser	TGC Cys	CAA Gln	GAG Glu	ACA Thr 345	GCT Ala	GTA Val	ACC Thr	CAG Gln	ACT Thr 350	1055
TAC Tyr	GGT Gly	GGC Gly	AAC Asn	TTA Leu 355	AAT Asn	GGA Gly	GAG Glu	CCA Pro	TGT Cys 360	GTC Val	TTA Leu	CCA Pro	TTC Phe	ACC Thr 365	TAC Tyr	1103
AAT Asn	GGC Gly	AGG Arg	ACG Thr 370	TTC Phe	TAC Tyr	TCC Ser	TGC Cys	ACC Thr 375	ACG Thr	GAA Glu	GGG Gly	CGA Arg	CAG G1n 380	GAC Asp	GGA Gly	1151
CAT His	Leu	TGG Trp 385	TGC Cys	AGC Ser	ACA Thr	Thr	TCG Ser 390	AAT Asn	TAT Tyr	GAG Glu	CAG Gln	GAC Asp 395	CAG Gln	AAA Lys	TAC Tyr	1199
TCT Ser	TTC Phe 400	TGC Cys	ACA Thr	GAC Asp	His	ACT Thr 405	GTT Val	TTG Leu	GTT Val	Gln	ACT Thr 410	CAA Gln	GGA Gly	GGA Gly	AAT Asn	1247
TCC / Ser / 415	AAT Asn	GGT Gly	GCC Ala	Leu	TGC Cys 420	CAC His	TTC Phe	CCC Pro	Phe	CTA Leu 425	TAC Tyr	AAC Asn	AAC Asn	CAC His	AAT Asn 430	1295

TAC Tyr	ACT Thr	GAT Asp	TGC Cys	ACT Thr 435	TCT Ser	GAG Glu	GGC Gly	AGA Arg	AGA Arg 440	GAC Asp	AAC Asn	ATG Met	AAG Lys	TGG Trp 445	TGT Cys	1343
GGG Gly	ACC Thr	ACA Thr	CAG Gln 450	AAC Asn	TAT Tyr	GAT Asp	GCC Ala	GAC Asp 455	CAG Gln	AAG Lys	TTT Phe	GGG Gly	TTC Phe 460	TGC Cys	CCC Pro	1391
ATG Met	GCT Ala	GCC Ala 465	CAC His	GAG Glu	GAA Glu	ATC Ile	TGC Cys 470	ACA Thr	ACC Thr	AAT Asn	GAA Glu	GGG Gly 475	GTC Val	ATG Met	TAC Tyr	1439
CGC Arg	ATT Ile 4 80	GGA Gly	GAT Asp	CAG Gln	TGG Trp	GAT Asp 485	AAG Lys	CAG Gln	CAT His	GAC Asp	ATG Met 490	GGT Gly	CAC His	ATG Met	ATG Met	1487
AGG Arg 495	TGC Cys	ACG Thr	TGT Cys	GTT Val	GGG Gly 500	AAT Asn	GGT Gly	CGT Arg	GGG Gly	GAA Glu 505	TGG Trp	ACA Thr	TGC Cys	ATT Ile	GCC Ala 510	1535
TAC Tyr	TCG Ser	CAA Gln	CTT Leu	CGA Arg 515	GAT Asp	CAG Gln	TGC Cys	ATT Ile	GTT Val 520	GAT Asp	GAC Asp	ATC Ile	ACT Thr	TAC Tyr 525	AAT Asn	1583
GTG Val	AAC Asn	GAC Asp	ACA Thr 530	TTC Phe	CAC His	AAG Lys	CGT Arg	CAT His 535	GAA Glu	GAG Glu	GGG Gly	CAC His	ATG Met 540	CTG Leu	AAC Asn	1631
TGT Cys	ACA Thr	TGC Cys 545	TTC Phe	GGT Gly	CAG Gln	GGT Gly	CGG Arg 550	GGC Gly	AGG Arg	TGG Trp	AAG Lys	TGT Cys 555	GAT Asp	CCC Pro	GTC Val	1679
GAC Asp	CAA Gln 560	TGC Cys	CAG Gln	GAT Asp	TCA Ser	GAG Glu 565	ACT Thr	GGG Gly	ACG Thr	TTT Phe	TAT Tyr 570	CAA Gln	ATT Ile	GGA Gly	GAT Asp	1727
TCA Ser 575	TGG Trp	GAG Glu	AAG Lys	TAT Tyr	GTG Val 580	CAT His	GGT Gly	GTC Val	AGA Arg	TAC Tyr 585	CAG Gln	TGC Cys	TAC Tyr	TGC Cys	TAT Tyr 590	1775
GGC Gly	CGT Arg	GGC Gly	ATT Ile	GGG Gly 595	GAG Glu	TGG Trp	CAT His	TGC Cys	CAA Gln 600	CCT Pro	TTA Leu	CAG Gln	ACC Thr	TAT Tyr 605	CCA Pro	1823
AGC Ser	TCA Ser	AGT Ser	GGT Gly 610	CCT Pro	GTC Val	GAA Glu	GTA Val	TTT Phe 615	ATC Ile	ACT Thr	GAG Glu	ACT Thr	CCG Pro 620	AGT Ser	CAG Gln	1871
CCC Pro	AAC Asn	TCC Ser 625	His	CCC Pro	ATC Ile	CAG Gln	TGG Trp 630	AAT Asn	GCA Ala	CCA Pro	CAG Gln	CCA Pro 635	Ser	CAC His	ATT Ile	1919

							GGC Gly		1967
							ATC Ile		2015
							ATC Ile		2063
		Glu					ACC Thr 700		2111
							ACT Thr		2159
							ACA Thr		2207
							TCG Ser		2255
							CAG Gln		2303
							CTG Leu 780		2351
							GAT Asp		2399
							GAT Asp		2447
							GTT Val		2495
							GTC Val		2543
							GAA G1 u 860		2591

AAC Asn	TCC Ser	GTC Val 865	ACC Thr	CTC Leu	AGT Ser	GAC Asp	TTG Leu 870	CAA Gln	CCT Pro	GGT Gly	GTT Val	CAG G1n 875	TAT Tyr	AAC Asn	ATC Ile	2639
ACT Thr	ATC Ile 880	TAT Tyr	GCT Ala	GTG Val	GAA Glu	GAA Glu 885	AAT Asn	CAA Gln	GAA Glu	AGT Ser	ACA Thr 890	CCT Pro	GTT Val	GTC Val	ATT Ile	2687
CAA Gln 895	CAA Gln	GAA Glu	ACC Thr	ACT Thr	GGC Gly 900	ACC Thr	CCA Pro	CGC Arg	TCA Ser	GAT Asp 905	ACA Thr	GTG Val	CCC Pro	TCT Ser	CCC Pro 910	2735
AGG Arg	GAC Asp	CTG Leu	CAG Gln	TTT Phe 915	GTG Val	GAA Glu	GTG Val	ACA Thr	GAC Asp 920	GTG Val	AAG Lys	GTC Val	ACC Thr	ATC Ile 925	ATG Met	2783
TGG Trp	ACA Thr	CCG Pro	CCT Pro 930	GAG Glu	AGT Ser	GCA Ala	GTG Val	ACC Thr 935	GGC Gly	TAC Tyr	CGT Arg	GTG Val	GAT Asp 940	GTG Val	ATC Ile	2831
											CTG Leu					2879
AAC Asn	ACC Thr 960	TTT Phe	GCA Ala	GAA Glu	GTC Val	ACC Thr 965	GGG Gly	CTG Leu	TCC Ser	CCT Pro	GGG Gly 970	GTC Val	ACC Thr	TAT Tyr	TAC Tyr	2927
TTC Phe 975	AAA Lys	GTC Val	TTT Phe	GCA Ala	GTG Val 980	AGC Ser	CAT His	GGG Gly	AGG Arg	GAG G1 u 985	AGC Ser	AAG Lys	CCT Pro	CTG Leu	ACT Thr 990	2975
GCT Ala	CAA Gln	CAG Gln	ACA Thr	ACC Thr 995	AAA Lys	CTG Leu	GAT Asp	GCT Ala	CCC Pro 100	Thr	AAC Asn	CTC Leu	CAG Gln	TTT Phe 100!	Val	3023
AAT Asn	GAA Glu	ACT Thr	GAT Asp 101	Ser	ACT Thr	GTC Val	CTG Leu	GTG Val 101	Arg	TGG Trp	ACT Thr	CCA Pro	CCT Pro 102	Arg	GCC Ala	3071
CAG Gln	ATA Ile	ACA Thr 102	Gly	TAC Tyr	CGA Arg	CTG Leu	ACC Thr 103	Val	GGC Gly	CTT Leu	ACC Thr	CGA Arg 103	Arg	GGC Gly	CAG Gln	3119
CCC Pro	AGG Arg 104	Gln	TAC Tyr	AAT Asn	GTG Val	GGT Gly 104	Pro	TCT Ser	GTC Val	TCC Ser	AAG Lys 105	Tyr	CCC Pro	CTG Leu	AGG Arg	3167
AAT Asn 105	Leu	CAG Gln	CCT Pro	GCA Ala	TCT Ser 106	Glu	TAC Tyr	ACC Thr	GTA Val	TCC Ser 106	CTC Leu 5	GTG Val	GCC Ala	ATA Ile	AAG Lys 1070	3215

GG G1	C AA(y Asr	C CAA	A GAG n Glu	AGC Ser 107	Pro	C AAA D Lys	GCC Ala	ACT Thr	GGA Gly 108	/ Val	Phe	C ACC	ACA Thr	CT(Lei 108	G CAG u Gln B5	3263
CC Pro	T GGG o Gly	G AGO Ser	TCT Ser 109	lle	CCA Pro	CCT Pro	TAC Tyr	AAC Asn 109	Thr	GAG Glu	GTG Val	ACT Thr	GAG Glu 110	Thr	ACC Thr	3311
AT(C GTG Val	ATC Ile 110	hhr	TGG Trp	ACG Thr	CCT Pro	GCT Ala III	Pro	AGA Arg	ATT	GGT Gly	TTT Phe 111	Lys	CTC Leu	G GGT Gly	3359
GT/ Val	CGA Arg 112	Pro	AGC Ser	CAG Gln	GGA Gly	GGA Gly 112	Glu	GCA Ala	CCA Pro	CGA Arg	GAA Glu 113	Val	ACT Thr	TCA Ser	GAC Asp	3407
TCA Ser 113	GGA Gly 5	AGC Ser	ATC Ile	GTT Val	GTG Val 114	Ser	GGC Gly	TTG Leu	ACT Thr	CCA Pro 114	Gly	GTA Val	GAA Glu	TAC Tyr	GTC Val 1150	3455
TAC Tyr	ACC Thr	ATC Ile	CAA Gln	GTC Val 115	Leu	AGA Arg	GAT Asp	GGA Gly	CAG Gln 1160	G1 u	AGA Arg	GAT Asp	GCG Ala	CCA Pro 116	Ile	3503
GTA Val	AAC Asn	AAA Lys	GTG Val 1170	Val	ACA Thr	CCA Pro	TTG Leu	TCT Ser 11 7 5	Pro	CCA Pro	ACA Thr	AAC Asn	TTG Leu 1180	His	CTG Leu	3551
GAG Glu	GCA Ala	AAC Asn 118	rro	GAC Asp	ACT Thr	GGA. Gly	GTG Val 1190	Leu	ACA Thr	GTC Val	TCC Ser	TGG Trp 1195	Glu	AGG Arg	AGC Ser	3599
ACC Thr	ACC Thr 1200	Pro	GAC Asp	ATT Ile	ACT Thr	GGT Gly 1205	Tyr	AGA Arg	ATT Ile	ACC Thr	ACA Thr 1210	Thr	CCT Pro	ACA Thr	AAC Asn	3647
GGC Gly 121	CAG Gln	CAG Gln	GGA Gly	Asn	TCT Ser 1220	Leu	GAA Glu	GAA Glu	Val	GTC Val 1225	His	GCT Ala	GAT Asp	CAG Gln	AGC Ser 1230	3695
TCC Ser	TGC Cys	ACT Thr	Phe	GAT Asp 1235	Asn	CTG . Leu	AGT Ser	Pro	GGC Gly 1240	Leu	GAG Glu	TAC Tyr	Asn	GTC Val 1245	Ser	3743
GTT Val	TAC Tyr	lhr	GTC Val 1250	AAG Lys	GAT Asp	GAC / Asp	Lys	GAA Glu 1255	AGT Ser	GTC Val	CCT Pro	Ile	TCT Ser 1260	Asp	ACC Thr	3791
ATC Ile	ATC Ile	CCA Pro 1265	GAG (Glu)	GTG (Val	CCC Pro	Gini	CTC / Leu 1270	ACT (Thr ,	GAC Asp	CTA Leu	Ser	TTT Phe 1275	GTT Val	GAT Asp	ATA Ile	3839
inr	GAT Asp 1280	ser .	AGC / Ser :	ATC (Gly	CTG / Leu / 1285	AGG T	TGG /	ACC (Thr I	Pro	CTA / Leu / 1290	AAC Asn	TCT Ser	TCC Ser	ACC Thr	3887

ATT ATT GGG T Ile Ile Gly T 1295	AC CGC ATC A yr Arg Ile T 1300	ACA GTA GTT [hr Va] Va]	GCG GCA GG Ala Ala Gl 1305	A GAA GGT A y Glu Gly I	TC CCT 3935 le Pro 1310
ATT TTT GAA G Ile Phe Glu A	AT TTT GTG T sp Phe Val T 1315	TAC TCC TCA Tyr Ser Ser	GTA GGA TA Val Gly Ty 1320	r Tyr Thr V	TC ACA 3983 al Thr 325
GGG CTG GAG C Gly Leu Glu P 1	CCG GGC ATT G Pro Gly Ile A 330	GAC TAT GAT Asp Tyr Asp 133	Ile Ser Va	T ATC ACT C al lle Thr L 1340	TC ATT 4031 eu Ile
AAT GGC GGC G Asn Gly Gly G 1345	AG AGT GCC (ilu Ser Ala F	CCT ACT ACA Pro Thr Thr 1350	CTG ACA CA Leu Thr G1	AA CAA ACG G In Gln Thr A 1355	CT GTT 4079 la Val
CCT CCT CCC A Pro Pro Pro T 1360	hr Asp Leu <i>A</i>	CGA TTC ACC Arg Phe Thr 1365	Asn Ile Gl	GT CCA GAC A Ly Pro Asp T 370	CC ATG 4127 hr Met
CGT GTC ACC T Arg Val Thr T 1375	GG GCT CCA C rp Ala Pro F 1380	CCC CCA TCC Pro Pro Ser	ATT GAT TT Ile Asp Le 1385	TA ACC AAC T eu Thr Asn P	TC CTG 4175 he Leu 1390
GTG CGT TAC T Val Arg Tyr S	TCA CCT GTG A Ser Pro Val 1 1395	AAA AAT GAG Lys Asn Glu	GAA GAT GT Glu Asp Va 1400	al Ala Glu L	TG TCA 4223 eu Ser 405
ATT TCT CCT T Ile Ser Pro S	TCA GAC AAT (Ser Asp Asn <i>H</i> 1410	GCA GTG GTC Ala Val Val 141	Leu Thr As	AT CTC CTG C sn Leu Leu P 1420	CT GGT 4271 ro Gly
ACA GAA TAT G Thr Glu Tyr V 1425	GTA GTG AGT (/al Val Ser \	GTC TCC AGT Val Ser Ser 1430	GTC TAC GA	AA CAA CAT G lu Gln His G 1435	AG AGC 4319 lu Ser
ACA CCT CTT A Thr Pro Leu A 1440	Arg Gly Arg (CAG AAA ACA Gln Lys Thr 1445	· Gly Leu As	AT TCC CCA A sp Ser Pro T 450	CT GGC 4367 hr Gly
ATT GAC TTT T Ile Asp Phe S 1455	TCT GAT ATT A Ser Asp Ile 1460	Thr Ala Asn	TCT TTT AC Ser Phe Th 1465	CT GTG CAC T hr Val His T	GG ATT 4415 rp Ile 1470
GCT CCT CGA G Ala Pro Arg A	GCC ACC ATC A Ala Thr Ile 1475	ACT GGC TAC Thr Gly Tyr	AGG ATC CO Arg Ile A 1480	rg His His P	CC GAG 4463 Pro Glu 485
CAC TTC AGT O	GGG AGA CCT Gly Arg Pro 1490	CGA GAA GAT Arg Glu Asp 149	Arg Val P	CC CAC TCT C ro His Ser A 1500	GG AAT 4511 arg Asn

TCC Ser	: ATO	C AC(Thi 15(r Le	C AC(u Thi	C AA(^ Asr	CTC Leu	ACT Thr 151	Pro	A GG(o Gl)	C ACA	A GAO	TAT 1 Tyr 151	^ Va	G GT 1 Va	C AGC 1 Ser	4559	}
ATC Ile	GT7 Val 152	Ala	r cr a Le	T AAT u Asr	GGC Gly	AGA Arg 152	Glu	GAA Glu	A AGT ı Ser	CCC Pro	TTA Leu 153	ιLeι	AT I I I	T GG e Gl	C CAA y Gln	4607	,
CAA Gln 153	Ser	ACA Thr	GT Va	T TCT 1 Ser	GAT Asp 154	Val	CCG Pro	AGG Arg	GAC Asp	CTG Leu 154	Glu	GTT Val	GT Val	r GC ⁻ I Ala	Γ GCG a Ala 1550	4655	
ACC Thr	CCC Pro	ACC Thr	Se)	C CTA Leu 155	Leu	ATC Ile	AGC Ser	TGG Trp	GAT Asp 156	Ala	CCT Pro	GCT Ala	GT(Val	ACA Thr 156		4703	
AGA Arg	TAT Tyr	TAC Tyr	AGO Arg 157	, lle	ACT Thr	TAC Tyr	GGA Gly	GAA Glu 157	Thr	GGA Gly	GGA Gly	AAT Asn	AGC Ser 158	Pro	GTC Val	4751	
CAG Gln	GAG Glu	TTC Phe 158	Inr	GTG Val	CCT Pro	GGG Gly	AGC Ser 1590	Lys	TCT Ser	ACA Thr	GCT Ala	ACC Thr 159	Ile	AGC Ser	GGC Gly	4799	
CTT Leu	AAA Lys 160	Pro	GGA Gly	GTT Val	GAT Asp	TAT Tyr 1605	Thr	ATC Ile	ACT Thr	GTG Val	TAT Tyr 1610	Ala	GTC Val	ACT Thr	GGC Gly	4847	
CGT Arg 1615	ыу	GAC Asp	AGC Ser	CCC Pro	GCA Ala 1620	Ser	AGC Ser	AAG Lys	CCA Pro	ATT Ile 1625	Ser	ATT Ile	AAT Asn	TAC Tyr	CGA Arg 1630	4895	
ACA Thr	GAA Glu	ATT Ile	GAC Asp	AAA Lys 1635	Pro	TCC Ser	CAG Gln	ATG Met	CAA Gln 1640	Val	ACC Thr	GAT Asp	GTT Val	CAG Gln 164	Asp	4943	
AAC Asn	AGC Ser	ATT Ile	AGT Ser 165	GTC Val O	AAG Lys	TGG Trp	Leu	CCT Pro 1 65 5	Ser	AGT Ser	TCC Ser	CCT Pro	GTT Val 1660	Thr	GGT Gly	4991	
TAC Tyr	AGA Arg	GTA Val 1665	1111	ACC Thr	ACT Thr	Pro	AAA Lys 1670	AAT Asn	GGA Gly	CCA Pro	GGA Gly	CCA Pro 1675	Thr	AAA Lys	ACT Thr	5039	
Lys	ACT Thr 1680	Ala	GGT Gly	CCA Pro	Asp	CAA Gln 1685	ACA (GAA Glu	ATG Met	Thr	ATT Ile 1690	Glu	GGC Gly	TTG Leu	CAG Gln	5087	
CCC / Pro 1695	ACA Thr	GTG Val	GAG Glu	TAT Tyr	GTG Val 1700	GTT /	AGT (Ser)	GTC Val	Tyr	GCT Ala 1705	CAG Gln	AAT Asn	CCA Pro	AGC Ser	GGA Gly 1710	5135	
SAG A	AGT Ser	CAG Gln	CCT Pro	CTG Leu 1715	GTT (Val (CAG /	ACT (Thr A	lla '	GTA Val 1720	ACC /	AAC . Asn	ATT Ile	GAT Asp	CGC Arg 1725	Pro	5183	

AAA GGA CTG Lys Gly Leu	GCA TTC ACT Ala Phe Thi 1730	GAT GTG Asp Val	GAT GTC Asp Val 1735	GAT TCC Asp Ser	ATC AAA / Ile Lys 1740	ATT GCT Ile Ala	5231
TGG GAA AGC Trp Glu Ser 174	CCA CAG GGG Pro Gln Gly 5	CAA GTT Gln Val 175	Ser Arg	TAC AGG Tyr Arg	GTG ACC Val Thr 1755	TAC TCG Tyr Ser	5279
AGC CCT GAG Ser Pro Glu 1760	GAT GGA ATO Asp Gly Ilo	CAT GAG His Glu 1765	CTA TTC Leu Phe	CCT GCA Pro Ala 177	Pro Asp	GGT GAA Gly Glu	5327
GAA GAC ACT Glu Asp Thr 1775	GCA GAG CTO Ala Glu Leo 178	ı Gln Gly	CTC AGA Leu Arg	CCG GGT Pro Gly 1785	TCT GAG Ser Glu	TAC ACA Tyr Thr 1790	5375
GTC AGT GTG Val Ser Val	GTT GCC TTO Val Ala Leo 1795	CAC GAT His Asp	GAT ATG Asp Met 1800	Glu Ser	Gln Pro	CTG ATT Leu Ile 1805	5423
GGA ACC CAG Gly Thr Gln	TCC ACA GC Ser Thr Al 1810	ATT CCT a lle Pro	GCA CCA Ala Pro 1815	ACT GAC Thr Asp	CTG AAG Leu Lys 1820	Phe Thr	5471
CAG GTC ACA Gln Val Thr 182	CCC ACA AG Pro Thr Se 5	C CTG AGC Leu Ser 183	Ala Gln	TGG ACA Trp Thr	CCA CCC Pro Pro 1835	AAT GTT Asn Val	5519
CAG CTC ACT Gln Leu Thr 1840	GGA TAT CG Gly Tyr Ar	A GTG CGG J Val Arg 1845	GTG ACC Val Thr	CCC AAG Pro Lys 185	Glu Lys	ACC GGA Thr Gly	5567
CCA ATG AAA Pro Met Lys 1855	GAA ATC AA Glu Ile As 18	n Leu Ala	CCT GAC Pro Asp	AGC TCA Ser Ser 1865	TCC GTG Ser Val	GTT GTA Val Val 1870	5615
TCA GGA CTT Ser Gly Leu	ATG GTG GC Met Val Al 1875	C ACC AAA a Thr Lys	TAT GAA Tyr Glu 1880	Val Ser	GTC TAT Val Tyr	GCT CTT Ala Leu 1885	5663
AAG GAC ACT Lys Asp Thr	TTG ACA AG Leu Thr Se 1890	C AGA CCA r Arg Pro	GCT CAG Ala Gln 1895	GGT GTT Gly Val	GTC ACC Val Thr 1900	Thr Leu	5711
GAG AAT GTO Glu Asn Val 190	AGC CCA CC Ser Pro Pr OS	A AGA AGG o Arg Arg 191	Ala Arg	GTG ACA Val Thr	GAT GCT Asp Ala 1915	ACT GAG Thr Glu	5759
ACC ACC ATO Thr Thr Ile 1920	ACC ATT AG Thr Ile Se	C TGG AGA r Trp Arg 1925	ACC AAG Thr Lys	ACT GAG Thr Glu 193	Thr Ile	ACT GGC Thr Gly	5807

TTC CAA GTT GAT GCC GTT CCA GCC AAT GGC CAG ACT CCA ATC CAG AGA Phe Gln Val Asp Ala Val Pro Ala Asn Gly Gln Thr Pro Ile Gln Arg 1935 1940 1945 1950	5855
ACC ATC AAG CCA GAT GTC AGA AGC TAC ACC ATC ACA GGT TTA CAA CCA Thr Ile Lys Pro Asp Val Arg Ser Tyr Thr Ile Thr Gly Leu Gln Pro 1955 1960 1965	5903
GGC ACT GAC TAC AAG ATC TAC CTG TAC ACC TTG AAT GAC AAT GCT CGG Gly Thr Asp Tyr Lys Ile Tyr Leu Tyr Thr Leu Asn Asp Asn Ala Arg 1970 1975 1980	5951
AGC TCC CCT GTG GTC ATC GAC GCC TCC ACT GCC ATT GAT GCA CCA TCC Ser Ser Pro Val Val Ile Asp Ala Ser Thr Ala Ile Asp Ala Pro Ser 1985 1990 1995	5999
AAC CTG CGT TTC CTG GCC ACC ACA CCC AAT TCC TTG CTG GTA TCA TGG Asn Leu Arg Phe Leu Ala Thr Thr Pro Asn Ser Leu Leu Val Ser Trp 2000 2005 2010	6047
CAG CCG CCA CGT GCC AGG ATT ACC GGC TAC ATC ATC AAG TAT GAG AAG Gln Pro Pro Arg Ala Arg Ile Thr Gly Tyr Ile Ile Lys Tyr Glu Lys 2015 2020 2025 2030	6095
CCT GGG TCT CCT CCC AGA GAA GTG GTC CCT CGG CCC CGC CCT GGT GTC Pro Gly Ser Pro Pro Arg Glu Val Val Pro Arg Pro Arg Pro Gly Val 2035 2040 2045	6143
ACA GAG GCT ACT ATT ACT GGC CTG GAA CCG GGA ACC GAA TAT ACA ATT Thr Glu Ala Thr Ile Thr Gly Leu Glu Pro Gly Thr Glu Tyr Thr Ile 2050 2055 2060	6191
TAT GTC ATT GCC CTG AAG AAT AAT CAG AAG AGC GAG CCC CTG ATT GGA Tyr Val Ile Ala Leu Lys Asn Asn Gln Lys Ser Glu Pro Leu Ile Gly 2065 2070 2075	6239
AGG AAA AAG ACA GAC GAG CTT CCC CAA CTG GTA ACC CTT CCA CAC CCC Arg Lys Lys Thr Asp Glu Leu Pro Gln Leu Val Thr Leu Pro His Pro 2080 2085 2090	6287
AAT CTT CAT GGA CCA GAG ATC TTG GAT GTT CCT TCC ACA GTT CAA AAG Asn Leu His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr Val Gln Lys 2095 2100 2105 2110	6335
ACC CCT TTC GTC ACC CAC CCT GGG TAT GAC ACT GGA AAT GGT ATT CAG Thr Pro Phe Val Thr His Pro Gly Tyr Asp Thr Gly Asn Gly Ile Gln 2115 2120 2125	6383
CTT CCT GGC ACT TCT GGT CAG CAA CCC AGT GTT GGG CAA CAA ATG ATC Leu Pro Gly Thr Ser Gly Gln Gln Pro Ser Val Gly Gln Gln Met Ile 2130 2135 2140	6431
TTT GAG GAA CAT GGT TTT AGG CGG ACC ACA CCG CCC ACA ACG GCC ACC Phe Glu Glu His Gly Phe Arg Arg Thr Thr Pro Pro Thr Thr Ala Thr 2145 2150 2155	6479

CCC ATA AGG CAT Pro Ile Arg His 2160	AGG CCA AGA CC Arg Pro Arg Pr 2165	A TAC CCG CCG o Tyr Pro Pro	AAT GTA GGA Asn Val Gly 2170	CAA GAA 6527 Gln Glu
GCT CTC TCT CAG Ala Leu Ser Gln 2175	ACA ACC ATC TC Thr Thr Ile Se 2180	A TGG GCC CCA r Trp Ala Pro 218	Phe Gln Asp	ACT TCT 6575 Thr Ser 2190
GAG TAC ATC ATT Glu Tyr Ile Ile	TCA TGT CAT CC Ser Cys His Pr 2195	T GTT GGC ACT o Val Gly Thr 2200	Asp Glu Glu	CCC TTA 6623 Pro Leu 2205
CAG TTC AGG GTT Gln Phe Arg Val 221	Pro Gly Thr Se	T ACC AGT GCC r Thr Ser Ala 2215	ACT CTG ACA Thr Leu Thr 2220	Gly Leu
ACC AGA GGT GCC Thr Arg Gly Ala 2225	Thr Tyr Asn Il	C ATA GTG GAG e Ile Val Glu 30	GCA CTG AAA Ala Leu Lys 2235	GAC CAG 6719 Asp Gln
CAG AGG CAT AAG Gln Arg His Lys 2240	GTT CGG GAA GA Val Arg Glu Gl 2245	G GTT GTT ACC u Val Val Thr	GTG GGC AAC Val Gly Asn 2250	TCT GTC 6767 Ser Val
AAC GAA GGC TTG Asn Glu Gly Leu 2255	AAC CAA CCT AC Asn Gln Pro Th 2260	G GAT GAC TCG ir Asp Asp Ser 226	Cys Phe Asp	CCC TAC 6815 Pro Tyr 2270
ACA GTT TCC CAT Thr Val Ser His	TAT GCC GTT GG Tyr Ala Val Gl 2275	GA GAT GAG TGG y Asp Glu Trp 2280	GAA CGA ATG Glu Arg Met	TCT GAA 6863 Ser Glu 2285
TCA GGC TTT AAA Ser Gly Phe Lys 229	Leu Leu Cys Gl	AG TGC TTA GGC n Cys Leu Gly 2295	TTT GGA AGT Phe Gly Ser 2300	Gly His
TTC AGA TGT GAT Phe Arg Cys Asp 2305	Ser Ser Arg Tr	GG TGC CAT GAC op Cys His Asp 310	AAT GGT GTG Asn Gly Val 2315	AAC TAC 6959 Asn Tyr
AAG ATT GGA GAG Lys Ile Gly Glu 2320	AAG TGG GAC CG Lys Trp Asp Ar 2325	GT CAG GGA GAA rg Gln Gly Glu	AAT GGC CAG Asn Gly Gln 2330	ATG ATG 7007 Met Met
AGC TGC ACA TGT Ser Cys Thr Cys 2335	CTT GGG AAC GG Leu Gly Asn G 2340	GA AAA GGA GAA Iy Lys Gly Glu 234	Phe Lys Cys	GAC CCT 7055 Asp Pro 2350
CAT GAG GCA ACC His Glu Ala Thr	TGT TAC GAT GA Cys Tyr Asp As 2355	AT GGG AAG ACA sp Gly Lys Thr 2360	TAC CAC GTA Tyr His Val	GGA GAA 7103 Gly Glu 2365

			AAG Lys 2370	Glu					Ile					Cys			7151
			CGG Arg 5					Asp					Pro				7199
		Ser	CCC Pro				Thr					Asn					7247
	Arg		CAT His			Thr					Asn						7295
			CCT Pro		Asp					Arg					Glu		7343
TAAA	TCAT	CT	TTCCA	ATCC	A GA	AGGAA	CAAG	CA1	GTCT	СТС	TGCC	CAAGA	ATC (CATCT	TAAACT	Γ	7403
GGAG	TGAT	GT 7	ragca	GACC	C A	CTTA	GAGT	TCT	тстт	тст	TTCT	TAAC	icc (CTTTG	стст	à	7463
GAGG	AAGT	TC 7	CCAG	CTTC	A GC	TCAA	CTCA	CAG	CTTC	тсс	AAGC	ATCA	vcc (CTGGG	AGTT	Γ	7523
сстс	AGGG	ו דד	гтстс	ATAA	A TO	AGGG	CTGC	ACA	TTGC	CTG	ттст	GCTT	CG A	AAGTA	TTCA	4	7583
TACC	GCTC	AG 7	TATTT	TAAA	T GA	AGTO	ATTO	TAA	GATT	TGG	TTTG	GGAT	CA A	ATAGO	AAAG		7643
ATAT	GCAG	icc A	ACCA	AGAT	G CA	AATG	TTTT	GAA	ATGA	TAT	GACC	CAAAA	TT 1	ГТААС	TAGG	4	7703
AAGT	CACC	CA A	ACAC	ттст	G CT	TTCA	CTTA	AGT	GTCT	GGC	CCGC	ATA	CT (STAGO	AACA/	4	7763
GCAT	GATO	TT G	STTAC	TGTG	A TA	TTTT	TAAAT	ATC	CACA	GTA							7803

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2446 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Arg Gly Pro Gly Pro Gly Leu Leu Leu Leu Ala Val Leu Cys 1 5 10 15

Leu Gly Thr Ala Val Pro Ser Thr Gly Ala Ser Lys Ser Lys Arg Gln
20 25 30

Ala	Gln	Gln 35	Met	Val	Gln	Pro	Gln 40	Ser	Pro	Val	A1a	Va1 45	Ser	Gln	Ser
Lys	Pro 50	Gly	Cys	Tyr	Asp	Asn 55	Gly	Lys	His	Tyr	G1n 60	Ile	Asn	Gln	G1n
Trp 65	G1 u	Arg	Thr	Tyr	Leu 70	Gly	Asn	Val	Leu	Val 75	Cys	Thr	Cys	Tyr	Gly 80
Gly	Ser	Arg	Gly	Phe 85	Asn	Cys	G1 u	Ser	Lys 90	Pro	G1 u	Ala	Glu	G1 u 95	Thr
Cys	Phe	Asp	Lys 100	Tyr	Thr	G1y	Asn	Thr 105	Tyr	Arg	Val	Gly	Asp 110	Thr	Tyr
Glu	Arg	Pro 115	Lys	Asp	Ser	Met	Ile 120	Trp	Asp	Cys	Thr	Cys 125	Ile	Gly	Ala
Gly	Arg 130	Gly	Arg	Ile	Ser	Cys 135	Thr	Ile	Ala	Asn	Arg 140	Cys	His	Glu	Gly
Gly 145	Gln	Ser	Tyr	Lys	Ile 150	Gly	Asp	Thr	Trp	Arg 155	Arg	Pro	His	Glu	Thr 160
Gly	Gly	Tyr	Met	Leu 165	Glu	Cys	Va1	Cys	Leu 170	Gly	Asn	Gly	Lys	Gly 175	Glu
Trp	Thr	Cys	Lys 180	Pro	Ile	Ala	Glu	Lys 185	Cys	Phe	Asp	His	Ala 190	Ala	G1y
Thr	Ser	Tyr 195	Val	Val	Gly	G1 u	Thr 200	Trp	Glu	Lys	Pro	Tyr 205	Gln	Gly	Trp
Met	Met 210	Val	Asp	Cys	Thr	Cys 215	Leu	Gly	Glu	Gly	Ser 220	Gly	Arg	Ile	Thr
Cys 225	Thr	Ser	Arg	Asn	Arg 230	Cys	Asn	Asp	Gln	Asp 235	Thr	Arg	Thr	Ser	Tyr 240
Arg	Ile	Gly	Asp	Thr 245	Trp	Ser	Lys	Lys	Asp 250	Asn	Arg	Gly	Asn	Leu 255	Leu
Gln	Cys	Ile	Cys 260	Thr	Gly	Asn	Gly	Arg 265	Gly	Glu	Trp	Lys	Cys 270	Glu	Arg
His	Thr	Ser 275	Va1	Gln	Thr	Thr	Ser 280	Ser	Gly	Ser	Gly	Pro 285	Phe	Thr	Asp
Val	Arg 290	Ala	Ala	Val	Tyr	G1n 295	Pro	Gln	Pro	His	Pro 300	Gln	Pro	Pro	Pro
Tyr 305	Gly	His	Cys	Val	Thr 310	Asp	Ser	Gly	Val	Val 315	Tyr	Ser	Val	Gly	Met 320

G1	n Tr	rp L	eu L	ys T 3	hr (25	Gln	G1	y As	n Ly	/s (G]n 330	Me	t Le	u Cy	/s T	hr C	ys 35	Leu
G1	y As	n G	1y V 3∙	al S 40	er (Cys	Gli	n Gl	u Th 34	ir <i>1</i> 15	41a	Va	1 Th	r Gl		nr T 50	yr	Gly
G1:	y As	n Le 35	eu A: 55	sn G	ly 0	llu	Pro	о Су 36	s Va O	17 L	.eu	Pr	o Ph	e Th 36	ir Ty 5	r A	sn	Gly
Arg	3 Th 37	r Ph O	ne Ty	yr S	er C	ys	Thr 375	Th	r Gl	u G	Пу	Arg	g G1 38	n As O	p G1	уН	is	Leu
50.	,		er Th		3	90						395)					400
			p Hi	40) 3					4	10					4]	15	
			u Cy 42	U					42	b					43	0		
		43.						440	1					445	5			
	730	,	n Ty			•	455						460	ı				Ala
403			u Gl		4 /	U						475					•	Ile 480
			ı Trı	40	5					49	90					49	5	
			61) 500	,					505						510)		
		313						520						525				
	J30		His			5	35						540					
545			Gln		221	U					5	555					5	60
			Ser	505	1					5/0	U					575		
			Val 580						585					Cys	590			
ıIУ	116	Gly 595	Glu	Trp	His	s Cy	ys (G1n 500	Pro	Lei	J G	ln		Tyr 605	Pro	Ser	S	er

Ser	Gly 610	Pro	Val	Glu	Val	Phe 615	Ile	Thr	Glu	ihr	Pro 620	Ser	Gin	Pro	Asn
Ser 625	His	Pro	Ile	Gln	Trp 630	Asn	Αla	Pro	Gln	Pro 635	Ser	His	Пе	Ser	Lys 640
Tyr	Ile	Leu	Arg	Trp 645	Arg	Pro	Lys	Asn	Ser 650	Val	Gly	Arg	Trp	Lys 655	G1 u
Ala	Thr	Ile	Pro 660	Gly	His	Leu	Asn	Ser 665	Tyr	Thr	Ile	Lys	Gly 670	Leu	Lys
Pro	Gly	Val 675	Val	Tyr	Glu	Gly	Gln 680	Leu	Ile	Ser	Ile	G1n 685	Gln	Tyr	Gly
His	Gln 690	Glu	Val	Thr	Arg	Phe 695	Asp	Phe	Thr	Thr	Thr 700	Ser	Thr	Ser	Thr
Pro 705	Val	Thr	Ser	Asn	Thr 710	Val	Thr	Gly	Glu	Thr 715	Thr	Pro	Phe	Ser	Pro 720
Leu	Val	Ala	Thr	Ser 725	Glu	Ser	Val	Thr	G1u 730	Ile	Thr	Ala	Ser	Ser 735	Phe
Val	Val	Ser	Trp 740	Val	Ser	Ala	Ser	Asp 745	Thr	Val	Ser	Gly	Phe 750	Arg	Val
Glu	Tyr	G1u 755	Leu	Ser	Glu	Glu	Gly 760	Asp	Glu	Pro	Gln	Tyr 765	Leu	Asp	Leu
Pro	Ser 770	Thr	Ala	Thr	Ser	Val 775	Asn	Ile	Pro	Asp	Leu 780	Leu	Pro	Gly	Arg
Lys 785	Tyr	Ile	Val	Asn	Va1 790	Tyr	Gln	Ile	Ser	Glu 795	Asp	Gly	Glu	Gln	Ser 800
Leu	Ile	Leu	Ser	Thr 805	Ser	Gln	Thr	Thr	Ala 810	Pro	Asp	Ala	Pro	Pro 815	Asp
Pro	Thr	Va1	Asp 820	Gln	Val	Asp	Asp	Thr 825	Ser	Ile	Val	Val	Arg 830	Trp	Ser
Arg	Pro	Gln 835	Ala	Pro	Ile	Thr	Gly 840	Tyr	Arg	Ile	Val	Tyr 845	Ser	Pro	Ser
Val	Glu 850	Gly	Ser	Ser	Thr	G1u 855	Leu	Asn	Leu	Pro	G1u 860	Thr	Ala	Asn	Ser
Val 865	Thr	Leu	Ser	Asp	Leu 870	Gln	Pro	Gly	Val	G1n 875	Tyr	Asn	Ile	Thr	11e 880
Tyr	Ala	Val	Glu	G1u 885	Asn	Gln	G1 u	Ser	Thr 890	Pro	Val	Val	Ile	Gln 895	Glr

Glu	Thr	Thr	Gly 900	Thr	Pro	Arg	Ser	Asp 905	Thr	Val	Pro	Ser	Pro 910	Arg	Asp
Leu	Gln	Phe 915	Val	Glu	Val	Thr	Asp 920	Va1	Lys	Val	Thr	Ile 925	Met	Trp	Thr
Pro	Pro 930	G1 u	Ser	Ala	Val	Thr 935	Gly	Tyr	Arg	Val	Asp 940	Val	Ile	Pro	Val
Asn 945	Leu	Pro	Gly	Glu	His 950	Gly	Gln	Arg	Leu	Pro 955	Ile	Ser	Arg	Asn	Thr 960
Phe	Ala	Glu	Val	Thr 965	Gly	Leu	Ser	Pro	Gly 970	Val	Thr	Tyr	Tyr	Phe 975	Lys
Val	Phe	Ala	Val 980	Ser	His	Gly	Arg	G1u 985	Ser	Lys	Pro	Leu	Thr 990	Ala	Gln
G1n	Thr	Thr 995	Lys	Leu	Asp	Ala	Pro 1000		Asn	Leu	Gln	Phe 1005		Asn	Glu
Thr	Asp 1010		Thr	Val	Leu	Val 1015		Trp	Thr	Pro	Pro 1 02 0		Ala	Gln	Ile
Thr 1025		Tyr	Arg	Leu	Thr 1030		Gly	Leu	Thr	Arg 1035		Gly	Gln	Pro	Arg 1040
	Tyr	Asn	Val	Gly 1045	Pro	Ser	Val	Ser	Lys 1050		Pro	Leu	Arg	Asn 1055	
Gln				1045 Glu					1050 Leu)				1055 Gly	5
Gln Gln	Pro	Ala	Ser 1060 Pro	1045 Glu)	5	Thr	Val	Ser 1065 Val	1050 Leu	Val	Ala	Ile	Lys 1070 Gln	1055 Gly)	Asn
Gln Gln Gln	Pro Glu	Ala Ser 1075	Ser 1060 Pro	1045 Glu) Lys	5 Tyr	Thr Thr	Val Gly 1080	Ser 1065 Val	1050 Leu S Phe	Val Thr	Ala Thr	Ile Leu 1085	Lys 1070 Gln	Gly Gly Pro	Asn Gly
Gln Gln Gln Ser	Pro Glu Ser 1090 Thr	Ala Ser 1075 Ile	Ser 1060 Pro Pro	Glu Glu Lys Pro	Tyr Ala	Thr Thr Asn 1095	Val Gly 1080 Thr	Ser 1065 Val) Glu	Leu Fhe	Val Thr Thr	Ala Thr Glu 1100 Lys	Ile Leu 1085 Thr	Lys 1070 Gln Thr	Gly Gly Pro	Asn Gly Val
Gln Gln Gln Ser Ile	Pro Glu Ser 1090	Ser 1075 Ile Trp	Ser 1060 Pro Pro	Glu Glu Lys Pro	Tyr Ala Tyr Ala 1110	Thr Thr Asn 1095 Pro	Val Gly 1080 Thr	Ser 1065 Val Glu Ile	Leu Phe Val	Val Thr Thr Phe 1115	Ala Thr Glu 1100 Lys	Ile Leu 1085 Thr)	Lys 1070 Gln Thr	Gly Pro Ile	Asn Gly Val Arg 1120 Gly
Gln Gln Ser Ile 1105	Pro Glu Ser 1090 Thr	Ser 1075 Ile Trp Gln	Ser 1060 Pro Pro Thr	Glu Lys Pro Gly 1125 Ser	Tyr Ala Tyr Ala 1110	Thr Asn 1095 Pro)	Val Gly 1080 Thr Arg	Ser 1069 Val Glu Ile Arg	Leu Phe Val Glu 1130	Val Thr Thr Phe 1115	Ala Thr Glu 1100 Lys	Ile Leu 1085 Thr Leu Ser	Lys 1070 Gln Thr Gly	Gly Pro Ile Val Ser 1135	Asn Gly Val Arg 1120 Gly
Gln Gln Ser Ile 1105 Pro	Pro Glu Ser 1090 Thr Ser	Ser 1075 Ile Trp Gln Val	Ser 1060 Pro Pro Thr Gly Val 1140 Leu	Glu Lys Pro Gly 1125 Ser	Tyr Ala Tyr Ala 1110 Glu	Thr Thr Asn 1095 Pro Ala Leu	Val Gly 1080 Thr Arg Pro	Ser 1065 Val Glu Ile Arg Pro 1145	Leu Phe Val Glu 1130 Gly	Val Thr Phe 1115 Val Val	Ala Thr Glu 1100 Lys Thr	Ile Leu 1085 Thr Leu Ser	Lys 1070 Gln Thr Gly Asp Val 1150	Gly Pro Ile Val Ser 1135 Tyr	Asn Gly Val Arg 1120 Gly Thr

	sn 185		Asp	Thr	Gly	Val 1190		Thr	Val	Ser	Trp 1195		Arg	Ser	Thr	Thr 1200
Pr	^0	Asp	Ile	Thr	Gly 1205		Arg	Ile	Thr	Thr 1210		Pro	Thr	Asn	Gly 1215	
G٦	n	Gly	Asn	Ser 1220		Glu	Glu	Val	Val 1225		Ala	Asp	Gln	Ser 1230	Ser)	Cys
Tł	ır	Phe	Asp 1235		Leu	Ser	Pro	Gly 1240		Glu	Tyr	Asn	Val 1245		Val	Tyr
Tł	ır	Val 1250		Asp	Asp	Lys	Glu 1255		Val	Pro	Ile	Ser 1260		Thr	Ile	Ile
	o 265		Va1	Pro	Gln	Leu 1270		Asp	Leu	Ser	Phe 1275		Asp	Ile	Thr	Asp 1280
Se	er	Ser	Ile	Gly	Leu 1285		Trp	Thr	Pro	Leu 12 9 0		Ser	Ser	Thr	Ile 1295	
G1	lу	Tyr	Arg	Ile 1300		Va1	Val	Ala	Ala 1305		Glu	Gly	Ile	Pro 1310	lle)	Phe
G 7	lu	Asp	Phe 1315		Tyr	Ser	Ser	Val 1320		Tyr	Tyr	Thr	Val 1325		Gly	Leu
G]	lu	Pro 1330		Ile	Asp	Tyr	Asp 1335		Ser	Val	Пe	Thr 1340		Ile	Asn	Gly
	l y 345		Ser	Ala	Pro	Thr 1350		Leu	Thr	Gln	Gln 135		Ala	Val	Pro	Pro 1360
Pr	^o	Thr	Asp	Leu	Arg 1365		Thr	Asn	Ile	Gly 1370		Asp	Thr	Met	Arg 1375	
Tł	ır	Trp	Ala	Pro 1380		Pro	Ser	Ile	Asp 138		Thr	Asn	Phe	Leu 1390	Val	Arg
Ty	γr	Ser	Pro 1395		Lys	Asn	Glu	Glu 1400		Va1	Ala	G1 u	Leu 140	Ser 5	Ile	Ser
Pı	0	Ser 1410		Asn	Ala	Val	Val 1415		Thr	Asn	Leu	Leu 1420		Gly	Thr	Glu
	yr 125		Va1	Ser	Val	Ser 1430		Val	Tyr	Glu	Gln 143		Glu	Ser	Thr	Pro 1440
Le	eu	Arg	Gly	Arg	Gln 144		Thr	G1 y	Leu	Asp 1450		Pro	Thr	Gly	Ile 145	Asp 5
PI	he	Ser	Asp	Ile 1460		Ala	Asn	Ser	Phe 146	.Thr 5	Val	His	Trp	Ile 1470	Ala O	Pro

- Arg Ala Thr Ile Thr Gly Tyr Arg Ile Arg His His Pro Glu His Phe 1475 1480 1485
- Ser Gly Arg Pro Arg Glu Asp Arg Val Pro His Ser Arg Asn Ser Ile 1490 1495 1500
- Thr Leu Thr Asn Leu Thr Pro Gly Thr Glu Tyr Val Val Ser Ile Val 1505 1510 1515 1520
- Ala Leu Asn Gly Arg Glu Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser 1525 1530 1535
- Thr Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro
 1540 1550
- Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr 1555 1560 1565
- Tyr Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu 1570 1580
- Phe Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys 1585 1590 1595 1600
- Pro Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly 1605 1610 1615
- Asp Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu 1620 1630
- Ile Asp Lys Pro Ser Gln Met Gln Val Thr Asp Val Gln Asp Asn Ser 1635 1640 1645
- Ile Ser Val Lys Trp Leu Pro Ser Ser Ser Pro Val Thr Gly Tyr Arg 1650 1660
- Val Thr Thr Pro Lys Asn Gly Pro Gly Pro Thr Lys Thr 1665 1670 1675 1680
- Ala Gly Pro Asp Gln Thr Glu Met Thr Ile Glu Gly Leu Gln Pro Thr 1685 1690 1695
- Val Glu Tyr Val Val Ser Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser 1700 1705 1710
- Gln Pro Leu Val Gln Thr Ala Val Thr Asn Ile Asp Arg Pro Lys Gly 1715 1720 1725
- Leu Ala Phe Thr Asp Val Asp Val Asp Ser Ile Lys Ile Ala Trp Glu 1730 1740
- Ser Pro Gln Gly Gln Val Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro 1745 1750 1755 1760

- Glu Asp Gly Ile His Glu Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp 1765 1770 1775
- Thr Ala Glu Leu Gln Gly Leu Arg Pro Gly Ser Glu Tyr Thr Val Ser 1780 1785 1790
- Val Val Ala Leu His Asp Asp Met Glu Ser Gln Pro Leu Ile Gly Thr 1795 1800 1805
- Gln Ser Thr Ala Ile Pro Ala Pro Thr Asp Leu Lys Phe Thr Gln Val 1810 1815 1820
- Thr Pro Thr Ser Leu Ser Ala Gln Trp Thr Pro Pro Asn Val Gln Leu 1825 1830 1835 1840
- Thr Gly Tyr Arg Val Arg Val Thr Pro Lys Glu Lys Thr Gly Pro Met 1845 1850 1855
- Lys Glu Ile Asn Leu Ala Pro Asp Ser Ser Ser Val Val Ser Gly 1860 1865 1870
- Leu Met Val Ala Thr Lys Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp 1875 1880 1885
- Thr Leu Thr Ser Arg Pro Ala Gln Gly Val Val Thr Thr Leu Glu Asn 1890 1895 1900
- Val Ser Pro Pro Arg Arg Ala Arg Va! Thr Asp Ala Thr Glu Thr Thr 1905 1910 1915 1920
- Ile Thr Ile Ser Trp Arg Thr Lys Thr Glu Thr Ile Thr Gly Phe Gln 1925 1930 1935
- Val Asp Ala Val Pro Ala Asn Gly Gln Thr Pro Ile Gln Arg Thr Ile 1940 1945 1950
- Lys Pro Asp Val Arg Ser Tyr Thr Ile Thr Gly Leu Gln Pro Gly Thr 1955 1960 1965
- Asp Tyr Lys Ile Tyr Leu Tyr Thr Leu Asn Asp Asn Ala Arg Ser Ser 1970 1975 1980
- Pro Val Val Ile Asp Ala Ser Thr Ala Ile Asp Ala Pro Ser Asn Leu 1985 1990 1995 2000
- Arg Phe Leu Ala Thr Thr Pro Asn Ser Leu Leu Val Ser Trp Gln Pro 2005 2010 2015
- Pro Arg Ala Arg Ile Thr Gly Tyr Ile Ile Lys Tyr Glu Lys Pro Gly 2020 2025 2030
- Ser Pro Pro Arg Glu Val Val Pro Arg Pro Arg Pro Gly Val Thr Glu 2035 2040 2045

- Ala Thr Ile Thr Gly Leu Glu Pro Gly Thr Glu Tyr Thr Ile Tyr Val 2050 2055 2060
- Ile Ala Leu Lys Asn Asn Gln Lys Ser Glu Pro Leu Ile Gly Arg Lys 2065 2070 2075 2080
- Lys Thr Asp Glu Leu Pro Gln Leu Val Thr Leu Pro His Pro Asn Leu 2085 2090 2095
- His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr Val Gln Lys Thr Pro 2100 2105 2110
- Phe Val Thr His Pro Gly Tyr Asp Thr Gly Asn Gly Ile Gln Leu Pro 2115 2120 2125
- Gly Thr Ser Gly Gln Gln Pro Ser Val Gly Gln Gln Met Ile Phe Glu 2130 2135 2140
- Glu His Gly Phe Arg Arg Thr Thr Pro Pro Thr Thr Ala Thr Pro Ile 2145 2150 2155 2160
- Arg His Arg'Pro Arg Pro Tyr Pro Pro Asn Val Gly Gln Glu Ala Leu 2165 2170 2175
- Ser Gln Thr Thr Ile Ser Trp Ala Pro Phe Gln Asp Thr Ser Glu Tyr 2180 2185 2190
- Ile Ile Ser Cys His Pro Val Gly Thr Asp Glu Glu Pro Leu Gln Phe 2195 2200 2205
- Arg Val Pro Gly Thr Ser Thr Ser Ala Thr Leu Thr Gly Leu Thr Arg 2210 2215 2220
- Gly Ala Thr Tyr Asn Ile Ile Val Glu Ala Leu Lys Asp Gln Gln Arg 2225 2230 2235 2240
- His Lys Val Arg Glu Glu Val Val Thr Val Gly Asn Ser Val Asn Glu 2245 2250 2255
- Gly Leu Asn Gln Pro Thr Asp Asp Ser Cys Phe Asp Pro Tyr Thr Val 2260 2265 2270
- Ser His Tyr Ala Val Gly Asp Glu Trp Glu Arg Met Ser Glu Ser Gly 2275 2280 2285
- Phe Lys Leu Leu Cys Gln Cys Leu Gly Phe Gly Ser Gly His Phe Arg 2290 2295 2300
- Cys Asp Ser Ser Arg Trp Cys His Asp Asn Gly Val Asn Tyr Lys Ile 2305 2310 2315 2320
- Gly Glu Lys Trp Asp Arg Gln Gly Glu Asn Gly Gln Met Met Ser Cys 2325 2330 2335

Thr Cys Leu Gly Asn Gly Lys Gly Glu Phe Lys Cys Asp Pro His Glu 2340 2345 2350
Ala Thr Cys-Tyr Asp Asp Gly Lys Thr Tyr His Val Gly Glu Gln Trp 2355 2360 2365
Gln Lys Glu Tyr Leu Gly Ala Ile Cys Ser Cys Thr Cys Phe Gly Gly 2370 2380
Gln Arg Gly Trp Arg Cys Asp Asn Cys Arg Arg Pro Gly Gly Glu Pro 2385 2390 2395 2400
Ser Pro Glu Gly Thr Thr Gly Gln Ser Tyr Asn Gln Tyr Ser Gln Arg 2405 2410 2415
Tyr His Gln Arg Thr Asn Thr Asn Val Asn Cys Pro Ile Glu Cys Phe 2420 2425 2430
Met Pro Leu Asp Val Gln Ala Asp Arg Glu Asp Ser Arg Glu 2435 2440 2445
(2) INFORMATION FOR SEQ ID NO:3:
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2179 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 311962

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GTCTAGGAGC CAGCCCCACC CTTAGAAAAG ATG TTT TCC ATG AGG ATC GTC TGC Met Phe Ser Met Arg Ile Val Cys	54
CTA GTT CTA AGT GTG GTG GGC ACA GCA TGG ACT GCA GAT AGT GGT GAA Leu Val Leu Ser Val Val Gly Thr Ala Trp Thr Ala Asp Ser Gly Glu 10 15 20	102
GGT GAC TTT CTA GCT GAA GGA GGA GGC GTG CGT GGC CCA AGG GTT GTG Gly Asp Phe Leu Ala Glu Gly Gly Gly Val Arg Gly Pro Arg Val Val 25 30 35 40	150
GAA AGA CAT CAA TCT GCC TGC AAA GAT TCA GAC TGG CCC TTC TGC TCT Glu Arg His Gln Ser Ala Cys Lys Asp Ser Asp Trp Pro Phe Cys Ser	198

	TGG Trp 60							246
	GAA Glu							294
	CTA Leu							342
	AAT Asn							390
	GAT Asp							438
	GTC Val 140							486
	CAG Gln							534
	GAC Asp							582
	TTA Leu							630
	CTT Leu							678
	CAC His 220							726
	AAT Asn							774
	ACA Thr							822
	GAG Glu							870

													AGC Ser			918
TCT Ser	GGG G1y	AGC Ser	TCT Ser 300	GGA Gly	CCT Pro	GGA Gly	AGT Ser	ACT Thr 305	GGA Gly	AAC Asn	CGA Arg	AAC Asn	CCT Pro 310	GGG Gly	AGC Ser	966
TCT Ser	GGG Gly	ACT Thr 315	GGA Gly	GGG Gly	ACT Thr	GCA Ala	ACC Thr 320	TGG Trp	AAA Lys	CCT Pro	GGG Gly	AGC Ser 325	TCT Ser	GGA Gly	CCT Pro	1014
													GGA Gly			1062
GGA Gly 345	AAC Asn	CAA Gln	AAC Asn	CCT Pro	GGA Gly 350	AGT Ser	CCT Pro	AGA Arg	CCT Pro	GGT Gly 355	AGT Ser	ACC Thr	GGA Gly	ACC Thr	TGG Trp 360	1110
													ACC Thr			1158
AGC Ser	TCT Ser	GTA Val	TCT Ser 380	GGT Gly	AGT Ser	ACT Thr	GGA Gly	CAA Gln 385	TGG Trp	CAC His	TCT Ser	GAA Glu	TCT Ser 390	GGA Gly	AGT Ser	1206
													AAC Asn			1254
													CCA Pro			1302
AGG Arg 425	AGA Arg	GAG Glu	TAC Tyr	CAC His	ACA Thr 430	GAA Glu	AAA Lys	CTG Leu	GTC Val	ACT Thr 435	AAA Lys	GGA Gly	GAT Asp	AAA Lys	GAG Glu 440	1350
CTC Leu	AGG Arg	ACT Thr	GGT Gly	AAA Lys 445	GAG Glu	AAG Lys	GTC Val	ACC Thr	TCT Ser 450	GGT Gly	AGC Ser	ACA Thr	ACC Thr	ACC Thr 455	ACG Thr	1398
CGT Arg	CGT Arg	TCA Ser	TGC Cys 460	TCT Ser	AAA Lys	ACC Thr	GTT Val	ACT Thr 465	AAG Lys	ACT Thr	GTT Val	ATT Ile	GGT Gly 470	CCT Pro	GAT Asp	1446
GGT Gly	CAC His	AAA Lys 475	GAA Glu	GTT Val	ACC Thr	AAA Lys	GAA Glu 480	GTG Val	GTG Val	ACC Thr	TCC Ser	GAA Glu 485	GAT Asp	GGT Gly	TCT Ser	1494

														GGT Gly		1542
														TTC Phe		1590
														ATG Met 535		1638
														GGC Gly		1686
														GCT Ala		1734
														ACT Thr		1782
														AGC Ser		1830
														ACA Thr 615		1878
														ATC Ile		1926
		CCT Pro 635									TAGA	CTAA	GT T	ТАААТ	ATTTC	1979
TGCA	CAGT	GT T	CCCA	TGGC	c cc	TTGC	ATTT	ССТ	тстт	AAC	тстс	TGTT	AC A	CGTC	ATTGA	2039
AACT	ACAC	TT T	TTTG	GTCT	G TT	TTTG	TGCT	AGA	CTGT	AAG	TTCC	TTGG	igg d	CAGG	GCCTT	2099
TGTC	TGTC	TC A	TCTC	TGTA	т тс	CCAA	ATGC	CTA	ACAG	TAC	AGAG	CCAT	GA C	TCAA	TAAAT	2159
ACAT	GTTA	AA T	GGAT	GAAT	G											2179

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 643 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Phe Ser Met Arg Ile Val Cys Leu Val Leu Ser Val Val Gly Thr 15

Ala Trp Thr Ala Asp Ser Gly Glu Gly Asp Phe Leu Ala Glu Gly Gly 25

Gly Val Arg Gly Pro Arg Val Val Glu Arg His Gln Ser Ala Cys Lys 45

Asp Ser Asp Trp Pro Phe Cys Ser Asp Glu Asp Trp Asn Tyr Lys Cys 55

Pro Ser Gly Cys Arg Met Lys Gly Leu Ile Asp Glu Val Asn Gln Asp 65

Phe Thr Asn Arg Ile Asn Lys Leu Lys Asn Ser Leu Phe Glu Tyr Gln 95

Lys Asn Asn Lys Asp Ser His Ser Leu Thr Thr Asn Ile Met Glu Ile Leu Arg Gly Asp Phe Ser Ser Ala Asn Asn Arg Asp Asn Thr Tyr Asn 125

Arg Val Ser Glu Asp Leu Arg Ser Arg Ile Glu Val Leu Lys Arg Lys 130 135 140

Val Ile Glu Lys Val Gln His Ile Gln Leu Leu Gln Lys Asn Val Arg 145 150 155 160

Ala Gln Leu Val Asp Met Lys Arg Leu Glu Val Asp Ile Asp Ile Lys 165 170 175

Ile Arg Ser Cys Arg Gly Ser Trp Ser Arg Ala Leu Ala Arg Glu Val 180 185 190

Asp Leu Lys Asp Tyr Glu Asp Gln Gln Lys Gln Leu Glu Gln Val Ile 195 200 205

Ala Lys Asp Leu Leu Pro Ser Arg Asp Arg Gln His Leu Pro Leu Ile 210 215 220

Lys Met Lys Pro Val Pro Asp Leu Val Pro Gly Asn Phe Lys Ser Gln 225 230 235 240

Leu	GIn	Lys	Val	Pro 245	Pro	Glu	Trp	Lys	A1a 250	Leu	lhr	Asp	Met	255	Gin
Met	Arg	Met	Glu 260	Leu	G1u	Arg	Pro	Gly 265	Gly	Asn	Glu	Ile	Thr 270	Arg	G1 y
Gly	Ser	Thr 275	Ser	Tyr	Gly	Thr	Gly 280	Ser	G1 u	Thr	G1 u	Ser 285	Pro	Arg	Asn
Pro	Ser 290	Ser	Ala	Gly	Ser	Trp 295	Asn	Ser	Gly	Ser	Ser 300	Gly	Pro	Gly	Ser
Thr 305	Gly	Asn	Arg	Asn	Pro 310	Gly	Ser	Ser	Gly	Thr 315	Gly	Gly	Thr	Ala	Thr 320
Trp	Lys	Pro	Gly	Ser 325	Ser	Gly	Pro	Gly	Ser 330	Ala	Gly	Ser	Trp	Asn 335	Ser
Gly	Ser	Ser	Gly 340	Thr	Gly	Ser	Thr	Gly 345	Asn	Gln	Asn	Pro	Gly 350	Ser	Pro
Arg	Pro	Gly 355	Ser	Thr	Gly	Thr	Trp 360	Asn	Pro	Gly	Ser	Ser 365	Glu	Arg	Gly
Ser	A1a 370	Gly	His	Trp	Thr	Ser 375	Glu	Ser	Ser	Val	Ser 380	Gly	Ser	Thr	Gly
Gln 385	Trp	His	Ser	Glu	Ser 390	Gly	Ser	Phe	Arg	Pro 395	Asp	Ser	Pro	Gly	Ser 400
Gly	Asn	Ala	Arg	Pro 405	Asn	Asn	Pro	Asp	Trp 410	Gly	Thr	Phe	Glu.	Glu 415	Val
Ser	Gly	Asn	Val 420	Ser	Pro	Gly	Thr	Arg 425	Arg	Glu	Tyr	His	Thr 430	Glu	Lys
Leu	Val	Thr 435	Lys	Gly	Asp	Lys	G1u 440	Leu	Arg -	Thr	Gly	Lys 445	Glu	Lys	Val
Thr	Ser 45 0	Gly	Ser	Thr	Thr	Thr 455	Thr	Arg	Arg	Ser	Cys 460	Ser	Lys	Thr	Val
Thr 465	Lys	Thr	Val	Ile	Gly 470	Pro	Asp	Gly	His	Lys 475	Glu	Val	Thr	Lys	G1u 480
Val	Val	Thr	Ser	G1u 485	Asp	Gly	Ser	Asp	Cys 49 0	Pro	Glu	Ala	Met	Asp 495	Leu
Gly	Thr	Leu	Ser 500	Gly	Ile	Gly	Thr	Leu 505	Asp	Gly	Phe	Arg	His 510	Arg	His
Pro	Asp	Glu 515	Ala	Ala	Phe	Phe	Asp 520	Thr	Ala	Ser	Thr	Gly 525	Lys	Thr	Phe

Pro	530	Phe	Phe	Ser	Pro	Met 535	Leu	Gly	Giu	Phe	540	Ser	Glu	Ihr	Glu		
Ser 545	Arg	Gly	Ser	Glu	Ser 550	G1y	Ile	Phe	Thr	Asn 555	Thr	Lys	Glu	Ser	Ser 560		
Ser	His	His	Pro	Gly 565	Ile	Ala	Glu	Phe	Pro 570	Ser	Arg	Gly	Lys	Ser 575	Ser		
Ser	Tyr	Ser	Lys 580	Gln	Phe	Thr	Ser	Ser 585	Thr	Ser	Tyr	Asn	Arg 59 0	Gly	Asp		
Ser	Thr	Phe 595	G1u	Ser	Lys	Ser	Tyr 600	Lys	Met	Ala	Asp	G1u 605	Ala	Gly	Ser		
Glu	Ala 610	Asp	His	Glu	Gly	Thr 615	His	Ser	Thṛ	Lys	Arg 620	Gly	His	Ala	Lys		
Ser 625	Arg	Pro	Val	Arg	Gly 630	Ile	His	Thr	Ser	Pro 635	Leu	Gly	Lys	Pro	Ser 640		
Leu	Ser	Pro															
(2)	INF	ORMAT	TION	FOR	SEQ	ID N	10:5:	:									
	(i)	() () ()	C) S1	ENGTI PE: FRANC		027 b leic ESS:	ase acio sino	pain d	^s								
	(ix)		ATURE A) NA B) LO	AME/K			1013										
	(xi)) SEC)UENC	CE DE	ESCRI	PTIC)N: S	SEQ :	ED NO):5:			-				
	ATG (Met /															4	47
	ACA Thr															ġ	95

25

143

ACT GAT TCT ACT.GTC CTG GTG AGA TGG ACT CCA CCT CGG GCC CAG ATA Thr Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile 35

20

			GTG Val						191
			TCT Ser 70						239
			ACC Thr						287
			ACT Thr						335
			AAC Asn						383
			CCA Pro						431
			GCA Ala 150						479
			TTG Leu						527
			GGA Gly						575
			TCT Ser						623
			CTC Leu						671
			AGA Arg 230						719
			GAA Glu						767
			CCC Pro						815

														ATC Ile		863
														ACC Thr		911
														ATT Ile		959
														ATT Ile		1007
														GGG Gly 350		1055
														AAT Asn		1103
GGC Gly	GAG Glu	AGT Ser 370	GCC Ala	CCT Pro	ACT Thr	ACA Thr	CTG Leu 375	ACA Thr	CAA Gln	CAA Gln	ACG Thr	GCT Ala 380	GTT Val	CCT Pro	CCT Pro	1151
														CGT Arg		1199
														GTG Val		1247
TAC Tyr	TCA Ser	CCT Pro	GTG Val	AAA Lys 420	AAT Asn	GAG Glu	GAA Glu	GAT Asp	GTT Val 425	GCA Ala	GAG Glu	TTG Leu	TCA Ser	ATT Ile 430	TCT Ser	1295
CCT Pro	TCA Ser	GAC Asp	AAT Asn 435	GCA Ala	GTG Val	GTC Val	TTA Leu	ACA Thr 440	AAT Asn	CTC Leu	CTG Leu	CCT Pro	GGT Gly 445	ACA Thr	GAA Glu	1343
TAT Tyr	GTA Val	GTG Val 450	AGT Ser	GTC Val	TCC Ser	AGT Ser	GTC Val 455	TAC Tyr	GAA Glu	CAA Gln	CAT His	GAG Glu 460	AGC Ser	ACA Thr	CCT Pro	1391
CTT Leu	AGA Arg 465	GGA Gly	AGA Arg	CAG Gln	AAA Lys	ACA Thr 470	GGT Gly	CTT Leu	GAT Asp	TCC Ser	CCA Pro 475	ACT Thr	GGC Gly	ATT Ile	GAC Asp	1439

TTT Phe 480	TCT Ser	GAT Asp	ATT Ile	ACT Thr	GCC Ala 485	AAC Asn	TCT Ser	TTT Phe	ACT Thr	GTG Val 490	CAC His	TGG Trp	ATT Ile	GCT Ala	CCT Pro 495	1487
CGA Arg	GCC Ala	ACC Thr	ATC Ile	ACT Thr 500	GGC Gly	TAC Tyr	AGG Arg	ATC Ile	CGC Arg 50 5	CAT His	CAT His	CCC Pro	GAG Glu	CAC His 510	TTC Phe	1535
AGT Ser	GGG Gly	AGA Arg	CCT Pro 515	CGA Arg	GAA Glu	GAT Asp	CGG Arg	GTG Val 520	CCC Pro	CAC His	TCT Ser	CGG Arg	AAT Asn 525	TCC Ser	ATC Ile	1583
ACC Thr	CTC Leu	ACC Thr 530	AAC Asn	CTC Leu	ACT Thr	CCA Pro	GGC Gly 535	ACA Thr	GAG Glu	TAT Tyr	GTG Val	GTC Val 540	AGC Ser	ATC Ile	GTT Val	1631
GCT Ala	CTT Leu 545	AAT Asn	GGC Gly	AGA Arg	GAG Glu	GAA Glu 550	AGT Ser	CCC Pro	TTA Leu	TTG Leu	ATT Ile 555	GGC Gly	CAA Gln	CAA Gln	TCA Ser	1679
ACA Thr 560	GTT Val	TCT Ser	GAT Asp	GTT Val	CCG Pro 565	AGG Arg	GAC Asp	CTG Leu	GAA Glu	GTT Val 570	GTT Val	GCT Ala	GCG Ala	ACC Thr	CCC Pro 575	1727
ACC Thr	AGC Ser	CTA Leu	CTG Leu	ATC Ile 580	AGC Ser	TGG Trp	GAT Asp	GCT Ala	CCT Pro 585	GCT Ala	GTC Val	ACA Thr	GTG Val	AGA Arg 590	TAT Tyr	1775
TAC Tyr	AGG Arg	ATC Ile	ACT Thr 595	TAC Tyr	GGA Gly	GAA Glu	ACA Thr	GGA Gly 600	GGA Gly	AAT Asn	AGC Ser	CCT Pro	GTC Val 605	CAG Gln	GAG Glu	1823
TTC Phe	ACT Thr	GTG Val 610	CCT Pro	GGG Gly	AGC Ser	AAG Lys	TCT Ser 615	ACA Thr	GCT Ala	ACC Thr	ATC Ile	AGC Ser 620	GGC Gly	CTT Leu	AAA Lys	1871
CCT Pro	GGA Gly 625	GTT Val	GAT Asp	TAT Tyr	ACC Thr	ATC Ile 630	ACT Thr	GTG Val	TAT Tyr	GCT Ala	GTC Val 635	ACT Thr	GGC Gly	CGT Arg	GGA Gly	1919
GAC Asp 640	AGC Ser	CCC Pro	GCA Ala	AGC Ser	AGC Ser 645	AAG Lys	CCA Pro	ATT	TCC Ser	ATT Ile 650	AAT Asn	TAC Tyr	CGA Arg	ACA Thr	GAA Glu 655	1967
ATT Ile	GAC Asp	AAA Lys	CCA Pro	TCC Ser 660	CAG Gln	ATG Met	CAA Gln	GTG Val	ACC Thr 665	GAT Asp	GTT Val	CAG Gln	GAC Asp	AAC Asn 670	AGC Ser	2015
ATT Ile	AGT Ser	GTC Val	AAG Lys 675	TGG Trp	CTG Leu	CCT Pro	TCA Ser	AGT Ser 680	TCC Ser	CCT Pro	GTT Val	ACT Thr	GGT Gly 685	TAC Tyr	AGA Arg	2063
GTA Val	ACC Thr	ACC Thr 690	ACT Thr	CCC Pro	AAA Lys	AAT Asn	GGA Gly 695	CCA Pro	GGA Gly	CCA Pro	ACA Thr	AAA Lys 700	ACT Thr	AAA Lys	ACT Thr	2111

					ACA Thr											2159
GTG Val 720	GAG Glu	TAT Tyr	GTG Val	GTT Val	AGT Ser 725	GTC Val	TAT Tyr	GCT Ala	CAG Gln	AAT Asn 730	CCA Pro	AGC Ser	GGA Gly	GAG Glu	AGT Ser 735	2207
CAG Gln	CCT Pro	CTG Leu	GTT Val	CAG Gln 740	ACT Thr	GCA Ala	GTA Val	ACC Thr	AAC Asn 745	ATT Ile	GAT Asp	CGC Arg	CCT Pro	AAA Lys 750	GGA Gly	2255
					GTG Val											2303
AGC Ser	CCA Pro	CAG Gln 770	GGG Gly	CAA Gln	GTT Val	TCC Ser	AGG Arg 775	TAC Tyr	AGG Arg	GTG Val	ACC Thr	TAC Tyr 780	TCG Ser	AGC Ser	CCT Pro	2351
GAG Glu	GAT Asp 785	GGA Gly	ATC Ile	CAT His	GAG Glu	CTA Leu 790	TTC Phe	CCT Pro	GCA Ala	CCT Pro	GAT Asp 795	GGT Gly	GAA Glu	GAA Glu	GAC Asp	2399
ACT Thr 800	GCA Ala	GAG Glu	CTG Leu	CAA Gln	GGC Gly 805	CTC Leu	AGA Arg	CCG Pro	GGT Gly	TCT Ser 810	GAG Glu	TAC Tyr	ACA Thr	GTC Val	AGT Ser 815	2447
GTG Val	GTT Val	GCC Ala	TTG Leu	CAC His 820	GAT Asp	GAT Asp	ATG Met	GAG Glu	AGC Ser 825	CAG Gln	CCC Pro	CTG Leu	ATT Ile	GGA Gly 830	ACC Thr	2495
					CCT Pro											2543
ACA Thr	CCC Pro	ACA Thr 850	AGC Ser	CTG Leu	AGC Ser	GCC Ala	CAG Gln 855	TGG Trp	ACA Thr	CCA Pro	CCC Pro	AAT Asn 860	GTT Val	CAG Gln	CTC Leu	2591
ACT Thr	GGA Gly 865	TAT Tyr	CGA Arg	GTG Val	CGG Arg	GTG Val 870	ACC Thr	CCC Pro	AAG Lys	GAG Glu	AAG Lys 875	ACC Thr	GGA Gly	CCA Pro	ATG Met	2639
AAA Lys 880	GAA Glu	ATC Ile	AAC Asn	CTT Leu	GCT Ala 885	CCT Pro	GAC Asp	AGC Ser	TCA Ser	TCC Ser 890	GTG Val	GTT Val	GTA Val	TCA Ser	GGA Gly 895	2687
CTT Leu	ATG Met	GTG Val	GCC Ala	ACC Thr 900	AAA Lys	TAT Tyr	GAA Glu	GTG Val	AGT Ser 905	GTC Val	TAT Tyr	GCT Ala	CTT Leu	AAG Lys 910	GAC Asp	2735

	TTG Leu															2783
	AAT Asn															2831
	ACA Thr 945															2879
	GAG Glu															2927
ACG Thr	GAA Glu	AGC Ser	CCC Pro	AGG Arg 980	AAC Asn	CCT Pro	AGC Ser	AGT Ser	GCT Ala 985	GGA Gly	AGC Ser	TGG Trp	AAC Asn	TCT Ser 990	GGG Gly	2975
	TCT Ser								Arg					Ser		3023
	GGA Gly		Thr					Pro					Pro			3071
	GGA Gly 1025	Ser					Ser					Ser				3119
	AAC Asn O					Arg					Gly					3167
	AGC Ser				Gly					Trp					Ser	3215
	TCT Ser			Thr					Ser					Phe		3263
	GAT Asp		Pro					Ala					Pro			3311
	ACA Thr 1105	Phe					Gly					Gly				3359
	TAC Tyr)					Leu					Asp					3407

	GAG AAG GTC Glu Lys Val 1140					Arg	3455
TCA TGC TCT Ser Cys Ser	AAA ACC GTT Lys Thr Val 1155	Thr Lys T	ACT GTT AT Thr Val Il 1160	T GGT CCT e Gly Pro	GAT GGT Asp Gly 1165	CAC His	3503
	ACC AAA GAA Thr Lys Glu)				Ser Asp		3551
	ATG GAT TTA Met Asp Leu						3599
	CAT AGG CAC His Arg His 120	Pro Asp G	Glu Ala Al				3647
	AAA ACA TTC Lys Thr Phe 1220					Glu	3695
	GAG ACT GAG Glu Thr Glu 1235	Ser Arg G					3743
AAT ACA AAG Asn Thr Lys 1250	GAA TCC AGT Glu Ser Ser O	TCT CAT C Ser His H 1255	CAC CCT GG His Pro Gl	G ATA GCT y Ile Ala 1260	Glu Phe	CCT Pro	3791
TCC CGT GGT Ser Arg Gly 1265	AAA TCT TCA Lys Ser Ser	AGT TAC A Ser Tyr S 1270	AGC AAA CA Ser Lys Gl	A TTT ACT n Phe Thr 1275	AGT AGC Ser Ser	ACG Thr	3839
	AGA GGA GAC Arg Gly Asp 128	Ser Thr F	Phe Glu Se				3887
GCA GAT GAG Ala Asp Glu	GCC GGA AGT Ala Gly Ser 1300	GAA GCC G Glu Ala A	GAT CAT GA Asp His Gl 1305	A GGA ACA u Gly Thr	CAT AGC His Ser 131	Thr	3935
	CAT GCT AAA His Ala Lys 1315	Ser Arg F					3983
CCT TTG GGG Pro Leu Gly 1330	AAG CCT TCC Lys Pro Ser O	CTG TCC (Leu Ser F 1335	CCC TAGACT Pro	ÀAGT TAAA	TAT		4027

- (2) INFORMATION FOR SEQ ID NO:6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1336 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala Gln Gln
1 5 10 15

Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn Glu Thr 20 25 30

Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile Thr 35 40 45

Gly Tyr Arg Leu Thr Val Gly Leu Thr Arg Arg Gly Gln Pro Arg Gln
50 55 60

Tyr Asn Val Gly Pro Ser Val Ser Lys Tyr Pro Leu Arg Asn Leu Gln
65 70 75 80

Pro Ala Ser Glu Tyr Thr Val Ser Leu Val Ala Ile Lys Gly Asn Gln 85 90 95

Glu Ser Pro Lys Ala Thr Gly Val Phe Thr Thr Leu Gln Pro Gly Ser 100 105 110

Ser Ile Pro Pro Tyr Asn Thr Glu Val Thr Glu Thr Thr Ile Val Ile 115 120 125

Thr Trp Thr Pro Ala Pro Arg Ile Gly Phe Lys Leu Gly Val Arg Pro 130 135 140

Ser Gln Gly Glu Ala Pro Arg Glu Val Thr Ser Asp Ser Gly Ser 145 150 155 160

Ile Val Val Ser Gly Leu Thr Pro Gly Val Glu Tyr Val Tyr Thr Ile 165 170 175

Gln Val Leu Arg Asp Gly Gln Glu Arg Asp Ala Pro Ile Val Asn Lys 180 185 190

Val Val Thr Pro Leu Ser Pro Pro Thr Asn Leu His Leu Glu Ala Asn 195 200 205

Pro Asp Thr Gly Val Leu Thr Val Ser Trp Glu Arg Ser Thr Thr Pro 210 215 220

Asp Ile Thr Gly Tyr Arg Ile Thr Thr Thr Pro Thr Asn Gly Gln Gln 225 230 240

WO 94/16085 PCT/US93/12687

Gly	Asn	Ser	Leu	G1u 245	Glu	Val	Val	His	Ala 250	Asp	Gln	Ser	Ser	Cys 255	Thr
Phe	Asp	Asn	Leu 260	Ser	Pro	Gly	Leu	G1u 265	Tyr	Asn	Val	Ser	Val 270	Tyr	Thr
Val	Lys	Asp 275	Asp	Lys	Glu	Ser	Val 280	Pro	Ile	Ser	Asp	Thr 285	Ile	Ile	Pro
Glu	Va1 290	Pro	Gln	Leu	Thr	Asp 295	Leu	Ser	Phe	Val	Asp 300	Пе	Thr	Asp	Ser
Ser 305	Ile	Gly	Leu	Arg	Trp 310	Thr	Pro	Leu	Asn	Ser 315	Ser	Thr	Ile	Ile	Gly 320
Tyr	Arg	Ile	Thr	Val 325	Val	Ala	Ala	Gly	G1u 330	G1 ý	Ile	Pro	Ile	Phe 335	Glu
Asp	Phe	Val	Tyr 340	Ser	Ser	Val	Gly	Tyr 345	Tyr	Thr	Val	Thr	Gly 350	Leu	Glu
Pro	Gly	Ile 355	Asp	Tyr	Asp	Ile	Ser 360	Val	Ile	Thr	Leu	Ile 365	Asn	Gly	Gly
Glu	Ser 370	Ala	Pro	Thr	Thr	Leu 375	Thr	Gln	Gln	Thr	Ala 380	Val	Pro	Pro	Pro
Thr 385	Asp	Leu	Arg	Phe	Thr 390	Asn	Ile	Gly	Pro	Asp 395	Thr	Met	Arg	Val	Thr 400
Trp	Ala	Pro	Pro	Pro 405	Ser	Ile	Asp	Leu	Thr 410	Asn	Phe	Leu	Val	Arg 415	Tyr
Ser	Pro	Val	Lys 420	Asn	Glu	Glu	Asp	Val 425	Ala	Glu	Leu	Ser	Ile 430	Ser	Pro
Ser	Asp	Asn 435	Ala	Val	Val	Leu	Thr 440	Asn	Leu	Leu	Pro	Gly 445	Thr	Glu	Tyr
Val	Val 450	Ser	Val	Ser	Ser	Val 455	Tyr	Glu	Gln	His	G1u 460	Ser	Thr	Pro	Leu
Arg 465	Gly	Arg	Gln	Lys	Thr 470	Gly	Leu	Asp	Ser	Pro 475	Thr	Gly	Ile	Asp	Phe 480
Ser	Asp	Ile	Thr	Ala 485	Asn	Ser	Phe	Thr	Val 490	His	Trp	Ile	Ala	Pro 495	Arg
Ala	Thr	Ile	Thr 500	Gly	Tyr	Arg	Ile	Arg 505	His	His	Pro	Glu	His 510	Phe	Ser
Gly	Arg	Pro 515	Arg	Glu	Asp	Arg	Val 520	Pro	His	Ser	Arg	Asn 525	Ser	Ile	Thr

Leu	530	Asn	Leu	Inr	Pro	535	inr	GIU	ıyr	Vai	540	Ser	lle	Val	Ala
Leu 545	Asn	Gly	Arg	G1u	G1u 550	Ser	Pro	Leu	Leu	Ile 555	Gly	Gln	Gln	Ser	Thr 560
Val	Ser	Asp	Val	Pro 565	Arg	Asp	Leu	Glu	Val 570	Val	Ala	Ala	Thr	Pro 575	Thr
Ser	Leu	Leu	Ile 580	Ser	Trp	Asp	Ala	Pro 585	Αla	Val	Thr	Val	Arg 590	Tyr	Tyr
Arg	Ile	Thr 595	Tyr	Gly	Glu	Thr	Gly 600	Gly	Asn	Ser	Pro	Val 605	Gln	Glu	Phe
Thr	Val 610	Pro	Gly	Ser	Lys	Ser 615	Thr	Ala	Thr	Ile	Ser 620	Gly	Leu	Lys	Pro
Gly 625	Val	Asp	Tyr	Thr	Ile 630	Thr	Val	Tyr	Ala	Val 635	Thr	Gly	Arg	Gly	Asp 640
Ser	Pro	Ala	Ser	Ser 645	Lys	Pro	Ile	Ser	Ile 650	Asn	Tyr	Arg	Thr	Glu 655	Ile
Asp	Lys	Pro	Ser 660	Gln	Met	Gln	Val	Thr 665	Asp	Val	Gln	Asp	Asn 670	Ser	Ile
Ser	Val	Lys 675	Trp	Leu	Pro	Ser	Ser 680	Ser	Pro-	Val	Thr	Gly 685	Tyr	Arg	Val
Thr	Thr 690	Thr	Pro	Lys	Asn	Gly 695	Pro	Gly	Pro	Thr	Lys 7 0 0	Thr	Lys	Thr	Ala
Gly 705	Pro	Asp	Gln	Thr	Glu 710	Met	Thr	Ile	Glu	Gly 715	Leu	Gln	Pro	Thr	Val 720
Glu	Tyr	Val	Val	Ser 725	Val	Tyr	Ala	Gln	Asn 730	Pro	Ser	Gly	Glu	Ser 735	Gln
Pro	Leu	Val	Gln 740	Thr	Ala	Val	Thr	Asn 745	Ile	Asp	Arg	Pro	Lys 750	Gly	Leu
Ala	Phe	Thr 755	Asp	Val	Asp	Val	Asp 760	Ser	Ile	Lys	Ile	Ala 765	Trp	Glu	Ser
Pro	G1n 770	Gly	Gln	Val	Ser	Arg 775	Tyr	Arg	Val	Thr	Tyr 780	Ser	Ser	Pro	G1 u
Asp 785	Gly	Ilе	His	Glu	Leu 790	Phe	Pro	Ala	Pro	Asp 795	Gly	Glu	Glu	Asp	Thr 800
Ala	G1 u	Leu	Gln	Gly 805	Leu	Arg	Pro	Gly	Ser 810	G1 u	Tyr	Thr	Va1	Ser 815	Val

Val	Ala	Leu	His 820	Asp	Asp	Met	G1u	Ser 825	Gln	Pro	Leu	Ile	Gly 830	Thr	Gln
Ser	Thr	Ala 835	Ile	Pro	Ala	Pro	Thr 840	Asp	Leu	Lys	Phe	Thr 845	Gln	Val	Thr
Pro	Thr 850	Ser	Leu	Ser	Ala	G1n 855	Trp	Thr	Pro	Pro	Asn 86 0	Val	Gln	Leu	Thr
Gly 865	Tyr	Arg	Val	Arg	Va1 870	Thr	Pro	Lys	Glu	Lys 875	Thr	Gly	Pro	Met	Lys 880
Glu	Ile	Asn	Leu	Ala 885	Pro	Asp	Ser	Ser	Ser 890	Val	Val	Val	Ser	Gly 895	Leu
Met	Val	Ala	Thr 900	Lys	Tyr	Glu	Val	Ser 905	Va1	Tyr	Ala	Leu	Lys 910	Asp	Thr
Leu	Thr	Ser 915	Arg	Pro	Ala	Gln	Gly 920	Va1	Val	Thr	Thr	Leu 925	Glu	Gly	G1y
Asn	Phe 930	Lys	Ser	Gln	Leu	Gln 935	Lys	Val	Pro	Pro	Glu 940	Trp	Lys	Ala	Leu
Thr 945	Asp	Met	Pro	Gln	Met 950	Arg	Met	Glu	Leu	G1u 955	Arg	Pro	Gly	Gly	Asn 960
Glu	Ile	Thr	Arg	Gly 965	Gly	Ser	Thr	Ser	Tyr 970	Gly	Thr	G1y	Ser	G1u 975	Thr
Glu	Ser	Pro	Arg 980	Asn	Pro	Ser	Ser	Ala 985	Gly	Ser	Trp	Asn	Ser 990	Gly	Ser
Ser	Gly	Pro 995	Gly	Ser	Thr	Gly	Asn 1000		Asn	Pro	Gly	Ser 1005		Gly	Thr
Gly	Gly 1010		Ala	Thr		Lys 1015		Gly	Ser	Ser	Gly 1020		G1y	Ser	A1a
Gly 1025		Trp	Asn	Ser	Gly 1030		Ser	Gly	Thr	Gly 1035		Thr	Gly	Asn	Gln 1040
Asn	Pro	Gly	Ser	Pro 1049		Pro	Gly	Ser	Thr 1050		Thr	Trp	Asn	Pro 1055	
Ser	Ser	Glu	Arg 1060		Ser	Ala	Gly	His 1065		Thr	Ser	Glu	Ser 1070	Ser	Val
Ser	Gly	Ser 1075		Gly	Gln	Trp	His 1080		Glu	Ser	Gly	Ser 108		Arg	Pro
Asp	Ser 1090		Gly	Ser	Gly	Asn 1095		Arg	Pro	Asn	Asn 1100	Pro	Asp	Trp	Gly

Thr Phe Glu Glu Val	Ser Gly Asn Val	Ser Pro Gly Ihr	Arg Arg Glu
1105	1110	1115	1120

- Tyr His Thr Glu Lys Leu Val Thr Lys Gly Asp Lys Glu Leu Arg Thr 1125 1130 1135
- Gly Lys Glu Lys Val Thr Ser Gly Ser Thr Thr Thr Arg Arg Ser 1140 1145 1150
- Cys Ser Lys Thr Val Thr Lys Thr Val Ile Gly Pro Asp Gly His Lys
- Glu Val Thr Lys Glu Val Val Thr Ser Glu Asp Gly Ser Asp Cys Pro 1170 1175 1180
- Glu Ala Met Asp Leu Gly Thr Leu Ser Gly Ile Gly Thr Leu Asp Gly 1185 1190 1195 1200
- Phe Arg His Arg His Pro Asp Glu Ala Ala Phe Phe Asp Thr Ala Ser 1205 1210 1215
- Thr Gly Lys Thr Phe Pro Gly Phe Phe Ser Pro Met Leu Gly Glu Phe 1220 1225 1230
- Val Ser Glu Thr Glu Ser Arg Gly Ser Glu Ser Gly Ile Phe Thr Asn 1235 1240 1245
- Thr Lys Glu Ser Ser Ser His His Pro Gly Ile Ala Glu Phe Pro Ser 1250 1255 1260
- Arg Gly Lys Ser Ser Ser Tyr Ser Lys Gln Phe Thr Ser Ser Thr Ser 1265 1270 1275 1280
- Tyr Asn Arg Gly Asp Ser Thr Phe Glu Ser Lys Ser Tyr Lys Met Ala 1285 1290 1295
- Asp Glu Ala Gly Ser Glu Ala Asp His Glu Gly Thr His Ser Thr Lys 1300 1305 1310
- Arg Gly His Ala Lys Ser Arg Pro Val Arg Gly Ile His Thr Ser Pro 1315 1320 1325
- Leu Gly Lys Pro Ser Leu Ser Pro 1330 1335

(2) INFORMATION FOR SEQ ID NO:7:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC1551	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GATCCCCGGG GAGCTCCTCG AGGCATG	27
(2) INFORMATION FOR SEQ ID NO:8:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC1552	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
CCTCGAGGAG CTCCCCGGG	19
(2) INFORMATION FOR SEQ ID NO:9:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2052	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
AATTCACCAT GGCAGTGAGT	20

,	
(2) INFORMATION FOR SEQ ID NO:10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2053	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
CATGACTCAC TGCCATGGTG	2
(2) INFORMATION FOR SEQ ID NO:11:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2491	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
CTAGATTAGA ATGGGGCC	18
(2) INFORMATION FOR SEQ ID NO:12:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 10 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	

(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2493

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12: CCATTCTAAT

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(2) INFORMATION FOR SEQ ID NO. 13:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 88 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3521	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TCGACTTAAG GACACTTTGA CAAGCAGACC AGCTCAGGGT GTTGTCACCA CTCTGGAGGG	60
AGGAAATTTT AAGAGCCAGC TTCAGAAG	88
(2) INFORMATION FOR SEQ ID NO:14:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 88 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3522	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GTACCTTCTG AAGCTGGCTC TTAAAATTTC CTCCCTCCAG AGTGGTGACA ACACCCTGAG	60
CTGGTCTGCT TGTCAAAGTG TCCTTAAG	88

I Claim:

- 1. A hybrid protein comprising a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein.
- 2. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 3. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926.
- 4. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 5. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336.
- 6. A hybrid protein according to claim 1 comprising the amino acid sequence of Sequence ID Number 6 from alanine, amino acid number 2 to Proline, amino acid number 1336.
- 7. An isolated DNA molecule encoding a hybrid protein comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein.

- 8. A DNA molecule according to claim 7 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin, a collagen binding domain of fibronectin.
- 9. A DNA molecule according to claim 7 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 10. A DNA molecule according to claim 7 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid number 1 to glutamic acid, amino acid number 926.
- 11. A DNA molecule according to claim 7 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 12. A DNA molecule according to claim 7 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.
- 13. A DNA molecule according to claim 7 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 14. A DNA molecule according to claim 7 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.

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- 15. A DNA molecule according to claim 7 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 3 to nucleotide 4013.
- 16. A DNA construct comprising a DNA molecule encoding a hybrid protein, wherein said DNA molecule comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a crosslinking domain from a second protein, and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule.
- 17. A DNA construct according to claim 16 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 18. A DNA construct according to claim 16 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 19. A DNA construct according to claim 16 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid 1 to Glutamic acid, amino acid number 926.
- 20. A DNA construct according to claim 16 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen α chain.
- 21. A DNA construct according to claim 16 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.

- 22. A DNA construct according to claim 16 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 23. A DNA construct according to claim 16 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 1 to nucleotide 4013.
- 24. A DNA construct according to claim 16 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.
- 25. A host cell containing a DNA construct according to claim 16.
- 26. A method for producing a hybrid protein comprising culturing a host cell according to claim 25 under conditions promoting the expression of the first DNA segment.

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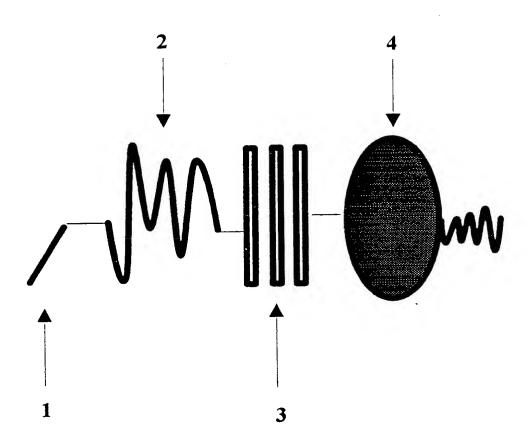


FIGURE 1

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Hybrid+FXIII+Thrombin

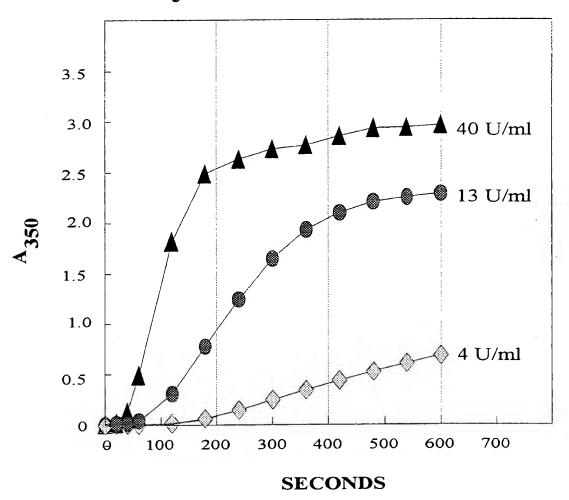


FIGURE 2

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Hybrid+FXIIIa

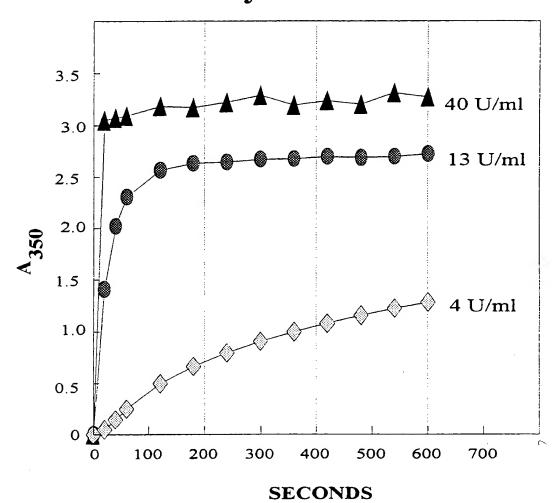


FIGURE 3

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FXIII+Thrombin

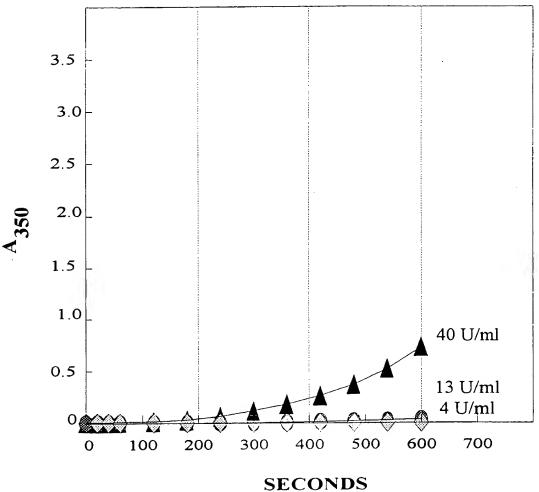


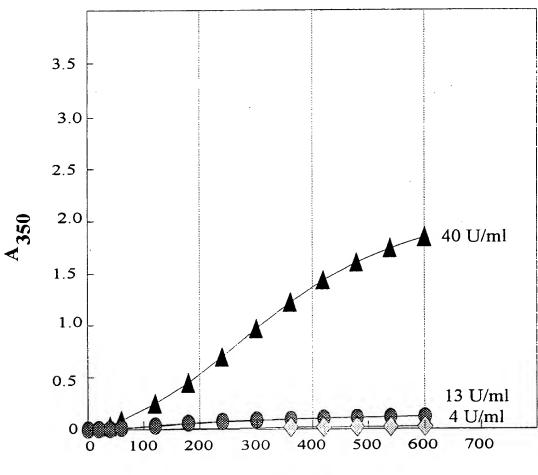
FIGURE 4

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FXIIIa



SECONDS

FIGURE 5